

EVALUATION OF DIFFERENT FUNGAL ANTAGONISTS, PLANT EXTRACTS AND OIL CAKES AGAINST FUSARIUM OXYSPORUM F.SP. CUMINI CAUSING WILT OF CUMIN. (CUMINUM CYMINUM L.)

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Trichoderma viride, T. harzianum, T. virens and T. hamatum isolated from rhizosphere of cumin significantly inhibited the mycelial growth of Fusarium oxysporum f.sp. cumini. Maximum inhibition of mycelial growth (77.77% and 75.55%) was recorded with local isolates of T. virens and T. viride respectively. Coiling of antagonists hyphae around the pathogen hyphae and finally lysis were observed. Out of five plant extracts and four oil cakes, plant extract of Datura stramonium, Calotropis procera and oil cakes of mustard and Sesamum, inhibited mycelial growth at the maximum in *in vitro* condition.

Key words : *Botanicals, cumin;* Fusarium oxysporum *f.sp.* cumini, Trichoderma harzianum, Trichoderma viride, Trichoderma hamatum, Trichoderma virens.

Cumin (Cuminum cyminum L.) is an important spice crop of Rajasthan. Wilt of cumin induced by Fusarium oxysporum f.sp. cumini is emerging as severe and destructive problem under favourable weather conditions during winter. The losses due to this disease is usually 5 to 80% (Mathur and Prasad, 1964a). Infection at seedling stage results in complete loss and in later infection either no seed is formed or it is thin, tiny and shrivelled. Reduction or elimination of soil borne inoculum is the only effective solution to overcome the problem and this may be achieved easily through fungal antagonists and soil amendments. Therefore studies were conducted to determine most effective fungal antagonists and soil amendments for further integration in the management practices of the disease.

MATERIALS AND METHODS

Leaf extracts of Phyllanthus amarus, Calotropis procera, Ricuinus communis, Datura stramonium,

Verbesena enceloides were tested for their fungicidal efficacy by the poisoned food technique (Nene and Thapliyal, 1993). The extraction from plant parts was done with the help of pestle and mortar. By adding equal amount of hot water, the extracted material was then filtered through muslin cloth, further 6ml of extract was mixed with 100 ml of molten PDA cooled to 45° C @1% (w/v) and sterilized in autoclave. Each treatment was replicated thrice with appropriate untreated controls. These were incubated for 3 days at $28\pm1^{\circ}$ C before recording radial mycelial growth of *Fusarium oxysporum* f. sp. *cumini.*

To determine the most effective soil amendment four oil cakes *viz.*, *Azadirachta indica*, *Gossypium* sp., *Sesamum indicum* and *Brassica campestris* were evaluated against pathogen in *in vitro*. Fine powdered oil cakes soaked in sterile water @ 1g/ml mixed in mixer and filtered (100% solution). Six ml of this solution was incorporated into 100ml PDA under sterilized condition in three replications. The radial growth of mycelium was recorded for each replication, separately.

RESULTS AND DISCUSSION

Effect of Antagonists : The results presented in Table-1 revealed that the isolate of *Trichoderma virens* and *Trichoderma viride* were the most effective in which maximum growth inhibition of mycelium of *Fusarium oxysporum* f.sp. *cumini* was recorded i.e. 77.7% and 75.55%. Next to it were isolates of *T. hamatum* and *T. harzianum* with their respective growth inhibition of 69.77% and 62.22%. *Trichoderma virens* grew over the colony of *Fusarium oxysporum* f.sp. *cumini* after 34 days of incubation. This overgrowth may be due to its fast growing nature, rapid sporulation, secretion of gliotoxin and / or cell wall lytic enzymes in dual culture. Thus it acts both as a competitor and antagonist. It is known to produce many enzymes i.e. chitinase, endochitinase, b1, 3 glucanase and secondary metabolites which possibly would have been involved in antagonism. T. virens covered the entire petriplate suppressing the growth of the pathogen. Fusarium could not be reisolated from this place, strongly suggesting suppression of the pathogen in dual culture. In case of other Trichoderma spp. host fungi initially grew towards the biocontrol agent, however the growing edge of *Fusarium* ceased before it could make contact with growing colony of Trichoderma. Later antagonist gradually overgrew the colony of Fusarium and inhibited the growth. Interaction with Trichoderma harzianum showed visible inhibition zone (2mm) while with T. viride.

Table-1 : Antagonistic effect of different fungi against *Eusarium oxysporum* f.sp. *cumini*

8 No	l est Mycoflora	Colony growth oxysport	Growth inhibition of Fusarium - oxysporum f.sp.	
		Control (mm)	Interaction (mm)	cumini (%)
I.	Trichoderma harziamm	45	17	62.22
2.	Trichoderma viride	45	11	75.55
	Trichoderma virens	45	10	77.77
4.	Trichoderma hamatum	45	13.6	69.77

Values are mean of 3 replicates

there was distinct change in colour at the zone of contact which become light yellowish. *T. hamatum* did not produce any demarcation zone. The growth inhibition of *Fusarium oxysporum* in presence of *Trichoderma* spp. could be attributed mainly due to antibiosis or hyperparasitism. Some substances were released by them like non-volatile sesquiterpene antibiotic. Hyphae of species of *Trichoderma* over powered the growth of pathogen and entered inside the mycelium of the pathogen. Sometimes the mycelium of pathogen was also found to be full of antagonist spores. Ultimately, host hyphae shrivelled and got killed. These observations indicated that pathogen lost their viability after colonization with antagonists (Elad *et al.*, 1990; Gokulapalan *et al.*, 1984; Lewis and Papavizas, 1980; Liu and Baker, 1980).

Effect of plant extracts : According to the results (table-2) plant leaf extracts of *Datura stramonium* and *Calotropis procera* were found to be highly significant in reducing the radial growth of the pathogen (72.92% and 68.75%) respectively. The other extracts in order of superiority were *Ricinus communis*, *Phyllanthus amarus* and *Verbesena enceloides*. The inhibitory effect of the plant extracts might be attributed to the presence of some antifungal toxicants. The aqueous extracts of these plants were found to affect the growth of the fungus.

It is therefore, encouraging to identify and characterize the active principle. Moreover, because of the water soluble nature of the toxic principle, it is ideal for developing into herbal pesticide. The presence of antifungal compounds in higher plants is well recognised (Mahadevan, 1982) and considered

Table-2 : Effect of plant extracts on radial growth of *Fusarium oxysporum* f.sp. *cumini*

S.No.	Plant extracts	Colony growth (mm) of Fusaruim oxysporum f.sp. cumini		Growth inhibition of <i>Fusarium</i>
		Control (mm)	Interaction (mm)	oxysporum f.sp. cumini (%)
1.	<i>Phyllanthus amarus</i> Hutch.	80	26.66	66.67
2	Calotropis procera (Ait) R.Br.	80	25	68.75
3.	<i>Ricinus communis</i> Linn.	80	25.33	68.33
4.	Datura stramonium Linn.	80	21.66	72.92
5.	<i>Verbesena enceloides</i> Benth & Hook	80	35	56.25

Values are mean of 3 replicates

valuable for plant disease control (Singh and Dwivedi, 1987). Various plant extracts have been evaluated for their antifungal property against different pathogens (Tripathi *et al.,* 2002; Sinha *et al.,* 2002; Mathur and Gurjar, 2002).

Effect of oil cakes : The results (Table 3) revealed that *Brassica campestris* oil cake inhibited the radial growth of the pathogen (73.07%) at the maximum followed by *Sesamum* oil cake (67.94%). Efficacy of different oil cakes was tested by many workers (Rajan,

1980; Rajan and Menon, 1970 and Ratnoo and Bhatnagar, 1983) in respect of reducing the propagules of the pathogen. The reduction in radial growth of the pathogen may be due to the presence of some inhibitory and antifungal substances.

It can be concluded from the present study that fungal antagonists viz. Trichoderma virens and Trichoderma viride, plant extracts of Datura stramonium and Calotropis procera and oil cakes of Brassica and Sesamum can be used in integrated management of Tusarium oxysporum f.sp. cumini infecting cumin plant at economical level in ecofriendly manner.

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S. No.	Oil cakes		Colony growth (mm) of Fusarium oxysporum f.sp. cumini		Growth inhibition of Fusarium
			Control (mm)	Interaction (mm)	= oxysporum f.sp. cumini (%)
L.	Sesamum (Sesamum ind	cake	78	25	67.94
2.	(Brassica (Brassica campestris)	cake	78	21	73.07
3.	Cotion	cake	78	30.33	61.11
4.	(Gossyphum s Neem (Azadirachta indica)	cake	78	26.33	66.24

Values are mean of 3 replicates

REFERENCES

Elad Y J Chet and J Katan 1990 *Trichoderma harzianum* a biocontrol agent effective against *Sclerotium rolfsii* and *Rhizoctonia solani*. Phytopathology, **70** : 119-121. Fokkema N J 1973 The role of saprophytic fungi in antagonism against *Drechslera sorokiniana* (*Helminthosporium sativum*) on agar plates and on rye leaves with pollen. Physiol. Pl Pathol **3** : 195-205.

Gokulapalan C and M C Nair 1984 Antagonism of a few fungi and bacteria against. *Rhizoctonia solani*. Indian J Microbiol **24** : 57-58.

Lewis J A and G C Papavizas 1980. Integrated control of *Rhizoctonia* fruit rot of cucumber Phytopathology

70:85-89.

Liu S and R Baker 1980. Mechanism of biological control in soil suppressive to *Rhizoctonia solani*. Phytopathology. **70** : 404-412.

Mahadevan A 1982 Biochemical aspects of plant disease resistance. Part I Performed inhibitory substances Prohibitions. Today and Tomorrows Printers and Publishers, New Delhi, India, 425 pp.

Mathur B L and N Prasad 1964a Studies on wilt disease of cumin caused by *Fusarium oxysporum* f.sp. *cumini*. Indian J. Agric. Sci. **34(2)** : 131-137.

Mathur Kamlesh R D, Singh and R B S, Gurjar 1995. Evaluation of different fungal antagonists, plant extracts and oil cakes against *Rhizoctonia solani* causing stem rot of chilli. Ann. Pl. Protec. Sci. **10(2)** : 319-322.

Nene Y L and P N Thapliyal 1993. Fungicides in plant disease control. Oxford and IBH Publ. Co. New Delhi, 507 pp.

Rajan K M 1980 Soil amendments in plant disease control. Int. Rice Newsl. 5 : 15.

Rajan K M and M R Menon 1970 Effect of organic soil amendment of plant growth and intensity of sheath blight of rice. Agril. Res. J. Kerala. **33** : 179-181.

Ratnoo R S and M K Bhatnagar 1993 Effect of straw oil cakes on ashy grey stem blight *Macrophomina phaseolina* (Jassi) Gold of cowpea Indian J. Mycol. Pl. Path. **23** : 186-188.

Singh R K and R S Dwivedi 1987 Effect of oil on *Sclerotium rolfsii,* causing root rot of barley. Indian Phytopath., **40** : 531-533.

Sinha A K, K P Verma, K C Agrawal and M P Thakur 2002 Antifungal activities of different plant extracts against *Colletotrichum capsici*. J. Mycol. Pl. Pathol. **32(2)** : 267 (Abstr).

Tripathi A K, K P Verma, K C Agrawal and S S Rao 2002 Effect of plant extracts on mycelial growth, sporulation and spore germination of *Alternaria lini* under *in vitro* condition J. Mycol. Pl. Pathol. **32(2)** : 268-269 (Abstr.)

Wood R K S 1951 The control of diseases of lettuce by use of antagonistic organisms. I. The control of *Botrytis cinerea* Pers. Annl. Appl. Biol. **38** : 203.