

IN-VITRO ANTIBACTERIAL ACTIVITY OF ANACYCLUS PYRETHRUM L. AND JASMINUM GRANDIFLORUM L.

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The petroleum ether, acetone, methanol and aqueous extracts of the root of *Anacyclus pyrethrum* and leaf of *Jasminum grandiflorum* were investigated for their antibacterial activity against the isolated and standard strains of *Staphylococcus aureus*, *Streptococcus mutans*, *S. sanguis*, *S. sobrinus*, *S. salivarius* and *Lactobacillus acidophilus* by using the agar well diffusion method. Amongst all the extracts tested, methanolic extract was comparably more effective to inhibit the growth of microbes than petroleum ether, acetone and aqueous extracts. The methanolic extract of *A. pyrethrum* and *J. grandiflorum* showed maximum antibacterial activity against *L. acidophilus* and *S. aureus* respectively. Phytochemical analysis of these extracts revealed the presence of antibacterial active agents such as alkaloids, flavonoids, glycosides, steroids, tannins, terpenoids, phenols and saponins.

Herbs are staging a comeback and herbal 'renaissance' is happening all over the globe. The herbal products today symbolize safety in contrast to the synthetics. Moreover, plants used in traditional medicine contain a vast array of substances that can be used to treat chronic and infectious diseases (Duraipandiyan et al. 2006). The present study was designed to evaluate A. pyrethrum (root) and J. grandiflorum (leaf) extracts against dental bacteria.

MATERIAL AND METHODS

Jasminum grandiflorum (chameli) were collected from the foot hills of Shivalik range of Himalayas in Hardwar and A. pyrethrum (akar-kara) was supplied by M/s Vijay Herbal Automation, Hardwar. These plants were further confirmed in the Department of Botany, Gurukula Kangri University, Hardwar and identified at Botanical Survey of India, Dehradun, Uttarakhand. The method of Alade and Irobi (1993) was adopted for the preparation of plant extracts with little

modification. The pathogenic organisms were selected for the study on the basis of their clinical pharmaceutical importance as well as for their potential to cause infection. The pathogens were isolated from the patients with dental diseases at Aggarwal dental clinic and identified in the Department of Botany and Microbiology, Gurukula Kangri University, Staphylococcus aureus MTCC Hardwar. 1144, Streptococcus mutans MTCC 890, S. sanguis ATCC 10556, S. sobrinus ATCC 33478. S. salivarius MTCC 1938 and Lactobacillus acidophilus MTCC 447 were procured from IMTECH, Chandigarh and National Chemical Lab. (NCL), Pune. The phytochemical analysis of plant extracts were carried out by using standard qualitative methods (Scalbert 1991, Chukwuran 1997). The antibacterial activity was performed by agar well diffusion method of Perez et al. (1990). The MIC was determined by dilution method using serially diluted two folds plant extracts according to Islam, et al. (2008.)

Table 1: *In-Vitro* antimicrobial activity of *A. pyrethrum* and *J. grandiflorum* indicated by diameter of inhibition zones (mm) at 100mg/ml.

S.No.	Pathogens	Anacyclus pyrethrum				Jasminum grandiflorum				Antibiotic
		Pt.ether	Acetone	Methanol	Aqueous	Pt.ether	Acetone	Methanol	Aqueous	Ofloxacin
1.	S. aureus	10±0.36	12±0.28	17±0.32	18±0.22	14±0.44	13±0.36	24±0.27	17±0.33	34±0.44
2.	S. aureus MTCC	9±0.34	14±0.31	18±0.30	16±0.30	9±0.22	10±0.32	14±0.20	15±0.28	34±0.44
3.	S. mutans	11±0.27	15±0.29	12±0.24	19±0.28	10±0.11	15±0.21	19±0.23	16±0.29	32±0.28
4.	S. mutans MTCC	10±0.14	24±0.20	18±0.25	17±0.34	12±0.28	14±0.24	20±0.28	14±0.31	32±0.30
5.	S. salivarius	9±0.20	12±0.26	14±0.30	15±0.30	11±0.24	15±0.27	14±0.24	19±0.34	30±0.31
6.	S.salivarius MTCC	10±0.25	14±0.31	17±0.32	13±0.34	14±0.32	16±0.30	16±0.22	20±0.22	31±0.28
7.	S. sanguis	9±0.37	13±0.32	15±0.30	17±0.36	10±0.30	12±0.31	15±	17±0.27	35±0.28
8.	S. sanguis ATCC	11±0.52	13±0.42	14±0.38	16±0.26	12±0.25	14±0.28	14±0.28	15±0.30	36±0.28
9.	S. sobrinus	10±0.37	14±0.40	12±0.35	19±0.44	11±0.32	13±0.30	15±0.34	18±0.30	28±0.27
10.	S. sobrinus ATCC	12±0.46	15±0.38	14±0.37	17±0.16	9±0.31	15±0.28	13±0.33	17±0.24	28±0.24
11.	L. acidophilus	10±0.40	16±0.35	14±0.40	18±0.28	12±0.41	14±0.35	15±0.33	16±0.24	30±0.25
12.	L.acidophilusMTCC	9±0.30	15±0.30	20±0.24	16±0.34	15±0.32	18±0.35	19±0.29	20±0.26	30±0.25

The data correspond to mean values \pm SE (n=3).

Results and Discussion: The present study showed significant antimicrobial activity of plant extracts against all the microorganisms (table-1). The methanol extract of A. pyrethrum showed the maximum antibacterial activity against the L. acidophilus MTCC (20mm) followed by petroleum ether, acetone and aqueous extract and minimum activity against isolated strain of Streptococcus sobrinus (12mm). In Jasminum grandiflorum the maximum antibacterial activity was shown by methanol extract against isolated strain of Staphylococcus aureus (24mm) followed by petroleum ether, acetone and aqueous extract and minimum activity against Streptococcus sobrinus ATCC (13mm). In general alcoholic extracts exhibited the highest degree of antibacterial activity as compared to aqueous extract (Ahmad et al., 1998). The Minimum Inhibition Concentration (MIC) value of A. pyrethrum was $\geq 8 \mu g/ml$ active against S. sanguis and the J. grandiflorum was $\geq 16 \,\mu\text{g/ml}$ active against S. aureus followed by Streptococcus mutans, S. sobrinus, S. salivarius and Lactobacillus acidophilus. The phytochemical screening of methanol extract

of plants showed that they contain alkaloids, flavonoids, glycosides, steroids, tannins, terpenoids, phenols and saponins. Tannins were absent in *A. pyrethrum* (roots) while amino acids, tannins and saponins were absent in *J. grandiflorum* (leaves). Plant compounds with antibacterial properties can be used as antibacterial agents in new drugs for the therapy of infectious diseases (table-2).

The *J. grandiflorum* is known to contain methyl anthranilate, benzyl alcohol

Table 2: Phytochemical analysis of the plants extracts in methanol.

S.No.	Phytochemical	Anacyclus	Jasminum		
		pyrethrum	grandiflorum.		
1.	Alkaloids	+	+		
2.	Flavonoids	+	+		
3.	Glycosides	+	+		
4.	Amino acid	+	-		
5.	Steroids	+	+		
6.	Tannins	-	-		
7.	Terpenoi ds	+	+		
8.	Saponins	+	-		

+: Present, -: Absent

and linalyl acetate. Our preliminary phytochemical study confirmed the presence of triterpenoids. The flavonoids (Villegas, 1997) and triterpenoids (Tsuchiya et al. 1996) are known to promote the wound- healing process mainly due to their astringent and antibacterial properties, which seems to be responsible for wound contraction and increased rate of epithelialisation. The anti-inflammatory action of triterpene alcohols present in the composite flowers have been well documented (Scortichini and Rossi 1991). The chloroform extract of the A. pyrethrum (root) at 1mg/plate produced inhibition of tobacco induced mutagenesis, using Ames Salmonella/ microsome assay. The extract also inhibited the nitrosation of methylurea in a dose dependent manner (Sukumaran and Kuttan 1992). The results support the traditional usage of the studied plants and suggests that the plant extracts possess compounds with antimicrobial properties that can be used as antibacterial agents in new drugs for the therapy of infectious diseases caused by pathogens. The most active extracts can be subjected to isolation of the therapeutic antibacterial compounds and carry out further pharmacological evaluation.

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REFERENCE:

Ahmad I Mehmood Z & Mohammad F 1998 Screening of some Indian medicinal plants for their antimicrobial properties. *Journal of Ethnopharmacology* **62** 183-193. Alade P I & Irobi O N 1993 Antimicrobial activity of crude leaf extracts of *Acalypha wilkesiana* **39** 171-174. Chukwurah B K C 1997 Antimicrobial activity of *Hollarrhena floridbunda* stem bark ethanol extracts. *Fitoterpia* **68** 180-181.

Duraipandiyan V, Ayyanar M & Ignacimuthu S 2006 Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from Tamil Nadu, India. *BMC Comp Alter Med* **6** 35-41.

Islam MA, Alam MM, Choudhury ME, Kobayashi N& Ahmed MU 2008 Determination of minimum inhibition concentration (MIC) of Cloxacillin for selected isolates of methicillin-resistant *Staphylococcus aureus* (MRSA) with their antibiogram. *Bangladesh Journal Veterinary Medicine* **6(1)** 121-126.

Perez C, Pauli M & Bazerque P 1990. An antibiotic assay by the well agar diffusion method. *Acta Biol Et Med Experimental* **15** 113-115.

Scalbert A 1991 Antimicrobial properties of tannins. *Phytochemistry* **30** 3875-3883.

Scortichini M & Rossi M P 1991 Preliminary in vitro evaluation of the antimicrobial activity of terpenes and terpenoids towards Erwinia amylovora (Burrill). Journal Applied Bacterial 71 109-112.

Sukumaran K & Kuttan R 1992 Inhibition of Tobacco-Induced mutagenesis by eugenol and plants extracts. *Amala Research Bulletin* **12** 96-101.

Tsuchiya H, Sato M, Miyazaki T, Fujiwara R, Tanigaki S & Ohyama M 1996 Comparative study on the antibacterial activity of phytochemical flavanones and methicillin-resistant *Staphylococcus aureus*. *Journal of Ethnopharmacology* **50** 27-34.

Vilegas L F, Fernandez I D, Maldonado H, Torres R Zavaleta A Vaisberg A J & Hammond G B 1997 Evaluation of the wound-healing activity of selected traditional medicinal plants from Peru. *Journal of Ethnopharmacology* **55** 193-200.