

NOVEL PLANT HORMONES- A REVIEW

¹POONAM MEENA, ²SAMPAT NEHRA AND ³P. C. TRIVEDI

¹Department of Botany, University of Rajasthan, Jaipur, India,²Department of Biotechnology, Birla Institute of Scientific Research, Statue Circle, Jaipur, Rajasthan, India, ³Vice Chancellor Jai Naran Yyas University, Jodhpur

Email DOi

Phytohormones are group of naturally occurring compounds secreted by plants which are synthesized at one place and translocated to another where they regulate the growth and differentiation, process of reproduction and even death. These phytohormones are required for appropriate stimulus when plants are subjected to stress or unwanted environment. Plant hormones, including auxin, cytokinin, abscisic acid, gibberellins, ethylene and recently evolved group of novel phytohormones including brassinosteroids, salicylic acid, jasmonates, polyamines, strigolactones and peptide hormones , significantly exhibit different functions in plant at cellular and molecular levels. The discovery of new class of phytohormones is new era of research in science to evaluate their various effect upon growth and development of plants.

Keywords: Brassinosteroids (BS), jasmonic acid(JA), peptide hormones(PH), polyamines(PA) and salicylic acid(SA)

Phytohormones are group of naturally occurring chemical compounds of plants required in small concentration and rarely effective at their synthesis, but regulate site of variety of physiological and morphological processes. Multiple hormones often influence a single process while single phytohormone can regulate many developmental and cellular processes at the same time. Novel Plant hormones, including brassinosteroids, salicylic acid, jasmonates, polyamines, strigolactones and peptide hormones beside conventional auxin, cytokinin, abscisic acid, gibberellins, ethylene phytohormones, are also able to regulate different functions in plants at cellular and molecular levels either individually or in crosstalk. Continuous effort has been made worldwide to explain their biosynthesis, mechanism of action, physiological role, interplay with other hormones and their potential of stress management. This article will review briefly about novel or sixth group of plant hormones with special consideration of brassinosteroids.

Salicylic acid/(SA): Raskin (1992) was introduced salicylic acid as a plant hormone. SA is a water soluble non-enzymatic antioxidant which acts as a plant growth regulator for different plant physiological processes like photosynthesis, shoot and root

development, flowering and senesence (Koo *et al.* 2020). SA combat abiotic stress (Sharma *et al.* 2020) and biotic stress (Ding & Ding 2020) respectively.

Jasmonic acid/(JA): Demole *et al.* (1962) isolated secondary metabolites as Jasmonic acid from the jasmine flower oils. Various developmental processes like vegetative growth, trichome development, anthocyanin biosynthesis, senescence and cell cycle regulation governed via JA (Gomi *et al.* 2020). Jasmonates have also been recognized as defence hormone which regulate various biotic and abiotic stress responses (Ruan *et al.* 2020 and Yang *et al.* 2019).

Polyamine/(PA): Polyamines (PA) are aliphatic amines, ubiquitous bioactive compounds occurring in microbes, plants, animal, and human semen and their main compounds are Putrescine (Put), spermidine (Spd) and spermines (spm), which exist mainly in free form inside higher plants. Since PA are fully protonated, they have been shown to bind to cell membrane and cell wall. PAs are key compounds in plant physiology (Takahashi 2020) such as for embryogenesis, organogenesis, senescence, fruit maturation and development. Polyamines played a significant role in biotic and abiotic stress (Sun *et al.* 2018, Chen *et al.* 2019 and Yu *et al.* 2019) management in plants.

Peptide Hormones/PH: Current research suggests that many secretory and nonsecretory peptide signals involved in multiple facets in growth and development of plant. Identification of these peptides involved biochemical purification method. Such peptide including, SYSTEMIN (defence), PHYTOSULPHOKINE/PSK (somatic embryogenesis), CLV3 (homeostasis or water stress), RAPID ALKALINIZATION FACTOR/RALF (arrestss growth of root), ENOD40 (nodule development), POLARIS/PLS (vascularization and expansion), INFLORESCENCE DEFICIENT IN ABSCISSION /IDA (organ abscission) etc. respectively (Matsabyashi and Sakagami 2006, Takahashi et al. 2018). Various peptides identified as membrane bound receptor kinase belonging to largest receptor like molecule significantly influence the plant family, growth and development (Gancheva 2019). These proteinases are involved in protein digestion of attacking pests (Hirakawa and Sawa 2019).

Strigolactone/(SL): Cook et al. (1966) first isolated strigol as a natural stimulant of Striga lutea, germination then these compounds were termed as strigolactones. SLs stimulates hyphal branching via AM symbiosis and support plant in uptake of available mineral nutrients like phosphate or in phosphate scarcity they produce higher amount of strigolactones (Mishra et al. 2017 and Bounmeester et al. 2019). SL promote seed germination in parasitic plants. It combat drought, salinity nutrition and temperature stress via AM symbiosis (Ruiz-Lozano, 2016) and it confers immunity against Pseudomonas syringae, Rhodococcus fascians and Pectobacterium carotovorum (Stes et al. 2015).

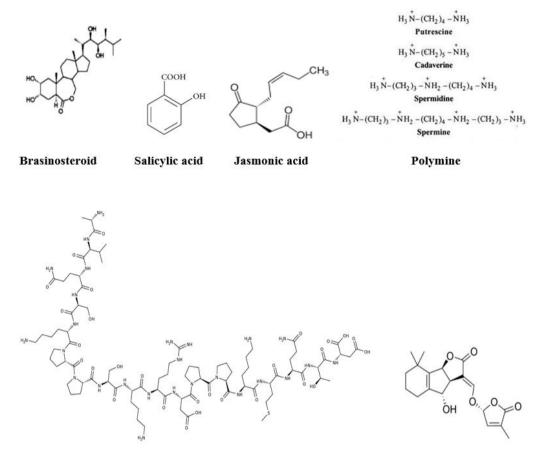
Brassinosteroids: Naturally occurring

brassinosteroids (BRs) are a polyhydroxy steroidal hormone, or known as sixth class of plant hormone. They have been isolated from many plants including fresh water and brown algae, monoplast, gymnosperm, dicots and monocots, (Fung and Sidall 1980). These hormones were explored when Mitchell et al. (1970) used organic extract of Brassica napus pollen treatment which induced cell division and stem elongation. Brassinolide is the most abundant and bioactive crystalline form and its structure determined by Grove et al. (1979) by X-ray crystallography. Individual species having variable distributions of BRs among different tissues like immature roots, seeds and flowers and pollens, have the large amount of BS upto 1-100 ng/g fw (fresh weight) whereas leaves and shoots contain lesser amounts 0.01–0.1 ng/g fw (Thompson et al. 1982). Brassinosteroids play significant contribution various disciplines of plant biology, in involving cell division cell differentiation and elongation, shoot and root growth, seed germination, stomatal movement photomorphogenesis, and fertility (Mitchell et al. 1941,1970 Laura et al. 2019 and Nolan et al. 2020) similar to animal hormones. BR have exclusive importance in mediating various abiotic stress like thermal, salinity, drought, freezing, metal toxicity and biotic disease stress. Extensive research upon brassinosteroids imply, that it has the potential enhance quality and quantity of to horticultural crops and save plants against biotic and abiotic stresses exiting in the local environment (Kang & Guo 2011).

Interplay between BRs and other plant hormones regulate a wide range of developmental and physiological processes. BRs depend on their crosstalk with other hormones like auxins and gibberellins for long distance effect. Exogenous application exhibit that excess BRs can be metabolize rapidly as plants synthesize them continuously to regulate their own growth and development.

Brassinosteroids (Brs) Cross Talk with

General structure of Novel hormones is given in Figure 1



Systemin

Strigolactone

Phytohormones:

BR and Auxin: BR and auxin are considered as potent phytohormones which significantly function as key regulators in different plant development processes such as hypocotyl elongation, stem elongation and root development (Zheng *et al.* 2019). Beside their coactive interplay, sometimes both interact antagonistically to control cell elongation, stem cell maintenance, gene expression control in root tips. Still balanced concentration of both hormones is needed for proper development of root in apple (Chaiwanon 2015, Zheng *et al.* 2019 & Ahmad *et al.* 2018).

BR and Gibberellin (GA): Clouse *et al.* (1996) reported, promising evidence of their interaction in between GA- and BR-deficient mutants of *Arabidopsis*, with the discovery of

the (being de-etiolated in the dark and dwarf stature in the light) remarkably resembled phenotypes. Crucial role of BR signifies genome expression in the GA regulation. The studies that indicated around 66.7% of brasinosteroid required for gene regulation of GA (Bai et al. 2012). GA promoted hypocotyl elongation was diminished in Arabidopsis with lower seedlings of biosynthesis of brassinosteroid, therefore both hormone are required for cell elongation (Gallego et al. 2012) while growth defective BR mutants can be rescue with GA (Peres et al. 2019).

BR and Cytokinin (CK): Cross talk between CKs and BRs associated with plant growth regulation. BR showed an prominent growth of lateral roots, thus their promising interplay

regulate growth and development of plant (Vercruyssen *et al.* 2003 & Ahmad *et al.*2018).

BR and Abscisic Acid (ABA):Up regulation of BIN2 and down regulation PP2C gene family mediates the BR signalling which is regulated by ABA (Zhang et al. 2009) causing decrement in BRs signalling pathway. Stomatal closure, root growth, seed germination, seedling formation significantly support the negative interplay between these hormones (Steber et al. 2001 & Takahashi et al. 2018). Usually defective BR signaling mutants (i.e, de-etiolated-2 mutant (det2), bin2-1, bri1, constitutive photo morphogenesis and dwarfism (cpd) posses high ABA sensitvity at the time of primary root formation, germination of seed and early seedling development, (Li et al. 2001, Nolan et al. 2017 & Fabregas et al. 2018).

BR and Ethylene (ET): Exogenous BR treated Arabidopsis seedlings exhibit increased ethylene biosynthesis level via an increase in ACS5 protein stability with its half life increment (Hensen et al. 2009) and ACS stability improved with high BRs which resist its degradation against 26S proteasome. Contrary to this, low concentration of BRs suppresses the ethylene biosynthesis with the enhanced level of BZR1/BES1 and main BR. Externally application of BR achieved good quality of fruiting, postharvest ripening via enhance ethylene production by increasing ACS2 and ACS4 genes transcription in Solanum lycopersicum (Zhu 2015, Fabregas et al. 2018 and Peres et al. 2019).

Physiological Effects of BR

Cell Elongation and differentiation: BR application at nM to μ M levels was effective in hypocotyl, peduncle and epicotyl, mesocotyl and coleoptile elongation (Clouse 1996). Endogenous BRs directly involved in cell expansion control in young vegetative tissue particularly while exogenous BR application were also found to be promising for plant growth with the xyloglucanase and expansin gene transcription, which promote cell wall loosening which in truns leads to stem elongation (Nolan *et al.* 2020).

Shoot and Root and leaves development: Remarkable increase in shoot and root growth was reported upon exogenous BR application (Hong *et al.* 2002). BRs regulate various physiological and developmental processes, including leaf growth as treatment of leaf mass and per unit leaf area of leaf (Sun *et al.* 2015). BR application was significantly increased during stress in some key processes like photosynthesis, carotenoid and chlorophyll content in cucumber (Li *et al.* 2016 & Nolan *et al.* 2020).

Pollen and Reproductive Biology: BR mutants have reduced fertility so brassinosteroids are also known as fertility hormone. Endogenous BR concentration is high inside pollen as it elongates the pollen tube suggested by *in vitro* studies (Hewitt 1985). Haploid production is favored by pollination initially and brassinolide treatment in *Arabidopsis thaliana* and *Brassica juncea*, induced haploid seed formation which later developed into stable plants (Kitani 1994).

Senescence: BRs accelerate senescence in normal plants evident with BR mutants which exhibit delayed senescence in *Arabidopsis* (Szekers *et al.* 1996). 24-epibrassinolide reduces malondialdehyde levels by oxidative degradation of lipids which was found to be a membrane protectant therefore remarkable delaying senescence observed (Ershova 1996 & Peres *et al.* 2019).

Brs Combat Stress in Plants: Human consumption of residual pesticides indirectly from vegetables and fruits gets reduced with an exogenous BRs application which display improved metabolism of pesticides and their removal in cucumbers. Research findings indicate that Brassinosteroids regulate the antioxidants and activities of antioxidative enzymes like glutathione reductase, catalase, guaiacol peroxidase, ascorbate peroxidase and superoxide dismutase to provide safeguard of plants under stress. Structural analogs of Brs have also been involved in the development, antioxidation, lipid peroxidation and protection. Application of BRS will reduce the oxidative stress in chickpea, soyabean, potato, rice and improved growth in plants recorded (Mazorra *et al.* 2002, Ozdemir *et al.* 2004, Almeida *et al.* 2005, Gruszka 2018 Peres *et al.* 2019, Nolan *et al.* 2020).

Biotic Stress: Plant resistance was induced by BRs, as exogenous application of BR to early field crop plants render immunity against various pathogenic infections (Yu et al. 2018). Exogenously BRs application enhance barley plants resistance against diverse pathogenic fungi (Ali et al. 2013). Phytophthora infection was significantly reduced in BRs treated seedlings of potato with the elevated level of phenolic, terpenoid compounds, ABA and ethylene level. BRs induced disease resistance mechanism is associated with the polyphenol metabolism via enhanced activities of polyphenoloxidase and peroxidase enzymes in cucumber plants. Virus infection reduced with BRS treated early plant seedlings (Rodkin et al. 1997) and yield increased upto 56%. Nakashita et al. (2003) reported that BRs treated plants remarkably reduced incidence of bacterial disease of Rice like bacterial blight (Xanthomonas orvzae) and rice blast (Magnaporthe grisea), respectively. Brassinolide treated egg masses of plant parasitic root knot nematode, Meloidogyne incognita significantly enhances hatching percentage (Ohri et al. 2002).

Abiotic Stress:

Salinity Stress: Sasse *et al.* (1995) reported 24-epibrassinolide treated seeds of *Eucalyptus camaldulensis*, under saline conditions resulted in good seed germination. BRs treated lettuce plants, have potential of ACC level reduction with ethylene production, consequently escape premature death and perfect protection effect to plants under

salinity (Mayank *et al.* 2004 & Peres *et al.* 2020). Saline medium containing rice seeds treated with BRs reinstate pigments considerably high rather than untreated plants under the same conditions (Anuradha and Rao 2003).

Thermal Stress: Chilling stress at 1-5° C was encounter with the 24-epibrassinolide application on rice, maizes pepper and cucumber (Rao et al. 2002 amd Anwar et al.2018) which significantly induced the photosynthesis rate, chlorophyll pigment, carbohydrate assimilation and regulated the gene expression to mitigate the required chilling stress. Increased ATP, Super oxide dismutase activity and proline level involved in the freezing tolerance in plants treated with BRs via osmoregulation and membrane stability (Li et al. 2017). Mitochondrial small heat shock proteins (MT-sHSP) expression was induced with the treatment of 24epibrassinolide upon tomato plants at 38° C or high temperature reported by Singh and Shono (2005) that imply thermotolerance has also achieved with application of Brs.

Drought Stress: Nilovskava et al. (2001) reported that BRs involved in improved resistance of plants under drought conditions and BRs treated such crops significantly increased crop yield value. 24-epibrassinolide solutions sprayed early seedlings were found to show their leaves to have large water content. Kagale et al. 2007 found that (A. thaliana and *B. napus* seedlings) 24epibrassinolide confers tolerance via altered expression of drought responsive genes under drought conditions in BRs treated plants have elevated level of stress responsive genes and membrane stability substances (osmoprotective metabolites) involved in antistress management process (Fàbregas et al. 2018).

Heavy Metal Toxicity Stress: Application of Brassinosteroids to different plants and crops renders the lowering of ingestion and accumulation of such heavy metal elements.

Treatment of 24-epibrassinolide reduced the absorption of radionuclides from cesium (Cs) and strontium (Sr) ions rich barley crop soil (Khripach et al. 1997). Epibrassinolide blocked the heavy metal (zinc, cadmium, lead and copper) accumulation in Chlorella vulgaris cells (Bajguz 2000) while mung bean (Phaseolus aureus) seedlings ameliorated Aluminium (Al) toxicity with the treatment with improved of brassinosteroids chlorophyll content and growth under Aluminium stress condition (Abdullahi et al. 2003). 28-Homobrassinolide treated nickel (Ni)-stressed B. juncea plants improved the seedling growth by increasing the activities of nitrate reductase and carbonic anhydrase photosynthesis rate of enzymes, and chlorophyll pigment (Alam et al. 2007). Ultimately BRs reciprocate the oxidative toxic metal stress via antioxidant enzyme introduction (Sharma et al. 2018).

CONCLUSION

Brassinosteroids are native, nonhazardous economic and ecofreindly compounds which significantly improve the crop productivity of unfertilized fields in lower concentration thus serve as potential tool for improvement of ecology and agriculture under extreme stressed environment. Latest technology focused upon stable synthetic BRs analogues formation and their genetic manipulatio which may become a suitable and effective strategy for larger yields of crop (Kang and Guo 2011). Since last many decades researcher constantly cover biochemical, structural, molecular and genomic approaches to elucidate mechanism brassinosteroid asction in growth, development, cross talk, biotic and abiotic stress response but despite such progress, there are still some unanswered puzzles exist in their mechanism or signaling. Advance technologies like Proteomics (Song et al. 2018) and genomics (Shahan 2019) via computational modeling may explore the genetic regulation of gene involved in induction and suppression of BRs mediated

response.

REFERENCES

Abdullahi BA, Gu X-G, Gan Q-L and Yang Y-H 2003. Brassinolide amelioration of aluminium toxicity in mung bean seedling growth. *J Plant Nutr* **26(9)** 1725-1735.

Ahmad F, Singh A and Kamal A 2018 Crosstalk of brassinosteroids with other phytohormones under various abiotic stresses J Appl Biol & B i o t e c h 6 (1) 5 6 - 6 2, D O I 10.7324/JABB.2018.60110.

Alam M M, Ali B and Ahmad A 2007 Effect of 28-homobrassinolide treatment on nickel toxicity in *Brassica juncea*. *Photosynthetica* **45(1)** 139-142.

Ali S S, Kumar G B S, Khan M and Doohan F M 2013 Brassinosteroid enhances resistance to Fusarium diseases of barley. *Phytopathology***103** 1260-1267.

Anwar A, Liu Y, Dong R, Bai L, Yu X and Li Y 2018 The physiological and molecular mechanism of brassinosteroid in response to stress: a review. *Biol. Research* **51** 46.

Almeida J M, Fidalgo F, Confraria A, Santos A, Pires H and Santos I 2005 Effect of hydrogen peroxide on catalase gene expression, isoform activities and levels in leaves of potato sprayed with homobrassinolide and ultrastructural changes in mesophyll cells. *Funct Plant Biol* **32** 707-720.

Anuradha S and Rao S S 2003 Application of brassinosteroids to rice seeds (*Oryza sativa* L.) reduced the impact of salt stress on growth, prevented photosynthetic pigment loss and increased nitrate reductase activity. *Plant Growth Regulation* **40(1)** 29-32.

Bai M Y, Shang J X, Oh E, Fan M, Bai Y, Zentella R, Sun T and Wang Z-Y 2012 Brassinosteroid, gibberellins and phytochrome Poonam Meena, Sampat Nehra and P. C. Trivedi

impinge on a common transcription module in *Arabidopsis*. *Nat Cell Biol* **14** 810–81.

Bajguz A 2007 Metabolism of brassinosteroids in plants *Plant Physiology and Biochem* **45(2)** 95–107.

Bounmeester H J, Fonne-Pfister R, Screpanti C and Mesmaeker AD 2019 Strigolactones: Plant Hormones with Promising Features. *J German Chem Society* https://doi.org/ 10.1002/ anie.201901626.

Chaiwanon J and Wang Z Y 2015 Spatiotemporal brassinosteroid signaling and antagonism with auxin pattern stem cell dynamics in Arabidopsis roots. *Curr Biol* **25** 1031–1042.

Chen D, Shao Q Yin L, Younis A and Zheng P 2019 Polyamine function in plants: Metabolism, Regulation on development and roles in abiotic stress response. *Front.* Plt Sci./http://doi.org/10.3389/fpls.2018.0945.

Clouse S D 1996 Molecular genetic studies confirm the role of brassinosteroids in plant growth and development. *Plant J* **10** 1–8.

Clouse S D, Langford M and McMorris T C 1996 A Brassinosteroid-Insensitive Mutant in *Arabidopsis thaliana* exhibits multiple defects in Growth and Development. *Plant Physiol* **111** 671–678.

Cook C E, Whichard L P, Turner B, Wall M E and Egley GH 1966. Germination of witchweed (*Striga lutea* Lour): isolation and properties of a potent stimulant. *Science* **154** 1189–1190. doi: 10.1120(asienee 154.2752.1180)

doi: 10.1126/science.154.3753.1189.

Demole E, Lederer E and Mercier D 1962 Isolement et détermination de la structure du jasmonate de méthyle, constituant odorant charactéristique de l'essence de jasmin. *Helvetica Chimica Acta*. **45** 675-685. J. Indian bot. Soc. Sp. Issue Vol. 100(A) 2020:120

Ding P and Ding Y 2020 Stories of SA: a plant defense hormone. *Trends in Plt Sci.* **25(6)** 549-565.

Ershova A and Khripach V 1996. Effect of epibrassinolide on lipid peroxidation in *Pisum sativum* at normal aeration and under oxygen deficiency. *Russ J Plant Physiol* 43 750–52.

Fàbregas N, Elena FL, Escamez DB, Tohge T, Martinez AC, Albacete A, Osorio S, Bustamante M, Riechmann JL, Yokota T, Conesa A, Fernie AR and Delgado AI 2018 Overexpression of the vascular brassinosteroid receptor BRL3 confers drought resistance without penalizing plant growth. *Nat Commun* **9** 4680.

Fung S and Siddall J B 1980_Stereoselective synthesis of brassinolide a plant growth promoting steroidal lactone *J Am Chem Soc* **102** 21 6580–6581.

Gallego-Bartolome J, Minguet E G, Grau-Enguix F, Abbas M, Locascio A, Thomas S G, Alabadi D and Blazquez MA 2012 Molecular mechanism for the interaction between gibberellin and brassinosteroid signalingpathways in Arabidopsis. *Proc Natl Acad Sci* USA, **109** 13446–13451.

Gancheva M S, Malovichko, Poliushkevich, Dodueva IE and Lutova LA 2019 Plant peptide hormones. *Russ Jon Plt Physiol.* **66** 171-189.

Gomi K 2020 Jasmonic acid -An essential plant hormone. *Internat. J. of Plt. Sci.* **21** 1261.

Grove MD, Spencer GF, Rohwedder WK, Mandava N, Worley JF, Warthen JD, Steffens GL, Flippen-Anderson JL and Cook JC 1979 Brassinolide, a plant growth-promoting steroid isolated from *Brassica napus* pollen". *Nature* **281** (5728) 216–217.

Gruszka D 2018 Crosstalk of the Brassinosteroid signalsone with phytohormonal and stress signaling components maintains a balance between the process of growth and Stress tolerance.

Hansen M, Chae H S and Kieber J J 2009 Regulation of ACS protein stability by cytokinin and brassinosteroid. *Plant J* **57** 606–614.

Hewitt F R, Hough T, O'Neill P, Sasse JM, Williams EG and Rowan KS 1985 Effect of brassinolide and other growth regulators on the germination and growth of pollen tubes of *Prunus avium* using a multiple hanging-drop assay. *Aust. J. Plant Physiol.* **12** 201–211.

Hirakawa Y and Sawa S 2019 Diverse function of plant peptide hormones in local signaling and development. *Curr Opinion Plt Biol* **51** 81-87.

Hong Z, Ueguchi-Tanaka M, Shimizu-Sato S, Inukai Y, Fujioka S, Shimada Y, Takatsuto S, Agetsuma M, Yoshida S and Watanabe Y 2002. Loss-of-function of a rice brassinosteroid biosynthetic enzyme, C-6 oxidase, prevents the organized arrangement and polar elongation of cells in the leaves and stem. *Plant J* **32** 495–508.

Kang YY and Guo SR 2011 Role of Brassinosteroids on Horticultural Crops. In Hayat S, Ahmad A (eds.). *Brassinosteroids A Class of Plant Hormone. Dordrecht, Netherlands Springer* 269–88.

Kagale S, Divi U K, Krochko J E, Keller W A and Krishna P (2007). Brassinosteroids confers tolerance in *Arabidopsis thaliana* and *Brassica napus* to a range of abiotic stresses. *Planta* **225** (2) 353-364.

Khripach V A, Zhabinskii V N, Litvinovskaya RP, Zavadskaya MI, Deeva VP and Vedeneev AN 1997 Preparation for the diminishing of radionuclides accumulation by plants and method of its application *Patent* BY 2806.

Kitani Y 1994. Induction of partheno-genetic

J. Indian bot. Soc. Sp. Issue Vol. 100(A) 2020:121

haploid plants with brassinolide. *Jpn J Genet* **69** 35–39.

Koo Y M, Heo A Y and Choi H W 2020 Salicylic Acid as a Safe Plant Protector and Growth Regulator. *Plant Pathol. J.* **36**(1) 1–10.

Laura A, Peres G L, Soares J, Rafael Tavares G, Righetto G, Zullo Marco A T, Mandava N B and Menossil M 2019. Brassinosteroids, the Sixth Class of Phytohormones A Molecular View from the Discovery to Hormonal Interactions in Plant Development and Stress Adaptation. *Int. J. Mol. Sci.* **1**-33.

Li H, Ye K, Shi Y, Cheng J, Zhang X and Yang S 2017. BZR1 positively regulates freezing tolerance via CBF-dependent and CBFindependent pathways in Arabidopsis. *Mol. Plant* **10** 545–559.

Li J, Nam KH, Vafeados D and Chory J 2001. BIN2, a new brassinosteroid-insensitive locus in *Arabidopsis*. *Plant Physiol* **127** 14–22.

Li J, Yang P, Kang J, Gan Y, Yu J, Calderón-Urrea A, Jian L, Zhang G, Feng Z and Xie J 2016 Transcriptome analysis of pepper revealed a role of 24-epibrassinolide in response to chilling. *Front Plant Sci* **7** 1–16.

Matsubayashi Y and Sakagami Y 2006 Peptide Hormones in plants. *Ann Rev of Plt Biol* **57** 6 4 9 - 6 7 4 . d o i : 10.1146/annurev.arplant.56.032604.144204.

Mayak S, Tirosh T and Glick B R 2004 Plant growth-promoting bacteria confer resistance in tomato plants to salt stress. *Plant Physiol. Biochem* **42** 565–572.

Mazorra LM, Nunez M, Hechavarria M, Coll F and Sanchez-Blanco MJ 2002 Infulence of brassinosteroids on antioxidant enzymes activity in tomato under different temperatures *Biol. Plant.* **45(4)** 593-596.

Mishra S, Upadhyay S and Shukla, RK 2017

Poonam Meena, Sampat Nehra and P. C. Trivedi

The Role of Strigolactones and Their Potential Cross-talk under Hostile Ecological Conditions in Plants. *Front. Physiol* 10 January 2017 | https://doi.org/10.3389/fphys.2016.00691.

Mitchell J W and Whitehead M R 1941 Responses of Vegetative parts of plants following application of extract of pollen from *Zea mays. Bot Gaz* **102** 770–791.

Mitchell J W, Mandava N B, Worley J F, Plimmer JR, Smith MV 1970 Brassins-A new family of planthormones from rape pollen. *Nature* **225** 1065–1066.

Nakashita H, Yasuda M, Nitta T, Asami T, Fujioka S, Arai Y, Sekimata K, Takatsuto S, Yamaguchi I and Yoshida S 2003 Brassinosteroids function in a broad range of disease resistance in tobacco and rice *PlJ*33(5) 887-898.

Nilovskaya N T, Ostapenko N V and Seregina II 2001. Effect of epibrassinolide on the productivity and drought resistance of spring wheat. *Agrokhimiya* **2** 46-50.

Nolan T, Chen J and Yanhai Y 2017 Cross-talk of Brassinosteroid signaling incontrolling growth and stress responses. *Biochemical J o u r n a l* **474**2641-2661, D O I 10.1042/BCJ20160633.

Nolan T M, Vukasinovic'N, Liu D, Russinova E and Yina Y 2020 Brassinosteroids Multidimensional Regulators of Plant Growth, Development, and Stress Responses (OPEN). *The Plant Cell* **32** 295 - 318.

Ohri P, Bhardwaj R and Khurma U R 2002 Role of brassinolide on juvenile emergence of *Meloidogyne incognita*. *Int J Nematol* **12** 175-178.

Ozdemir F, Bor M, Demiral T and Turkan I 2004 Effects of 24-epibrassinolide on seed germination, seedling growth, lipid peroxidation, proline content and antioxidative J. Indian bot. Soc. Sp. Issue Vol. 100(A) 2020:122

system of rice (*Oryza sativa* L.) under salinity stress. *Plant Growth Regul* **42** 203-211.

Peres A G L, Soares J S, Tavares R G, Rightetto G, Zullo MAT, Mandava B and Menossi M 2018 Brassinosteroids, the Sixth Class of Phytohormones A Molecular View from the Discovery to Hormonal Interactions in Plant Development and Stress Adaptation. *Int J Mol Sci* **20(2)** 331.

https//doi.org/10.3390/ijms20020331.

Raskin I 1992 Role of salicylic acid in plants. *Annu Rev Plant Physiol.* **43** 439–63.

Rao S S R, Vardhini B V V, Sujata E and Anuradha S 2002 Brassinosteroids – a new Class of Phytohormones. *Current Science* **82** 1239-1245.

Rodkin A I, Konovalova G I and Bobrick A O 1997. Efficiency of application of biologically active substances in primary breeding of potato. In *Plant Growth and Development Regulators. Moscow* 317-318.

Ruan J, Zhou Y, Zhou M, Yan Z, Khurshid M, Weng W, Cheng J and Zhang, K 2020 Jasmonic acid signaling pathways in plants. *Int J Mol Sci* **20** 2479.

Ruiz-Lozano J M, Aroca R, Zamarreño A M, Molina S, Andreo-Jiménez B and Porcel R 2016 Arbuscular mycorrhizal symbiosis induces strigolactone biosynthesis under drought and improves drought tolerance in lettuce and tomato. *Plant Cell Environ* **39** 441–452. doi: 10.1111/pce.12631

Sasse J M, Smith R and Hudson I 1995 Effect of 24-epibrassinolide on germination of seeds of *Eucalyptus camaldulensis* in saline conditions. *Proc Plant Growth Regul Soc Am* **22** 136-141.

Sharma A Sindhu G P S, Araniti F, Bali A S, Shahzad B, Tripathi DK, Brestic M and Landi M 2020 The Role of Salicylic Acid in Plants exposed to heavy met als. Molecules **25(3)** 540 https://doi.org/10.3390/molecules25030540

Sharma I, Sharma A, Pati P and Bhardwaj R 2018 Brassinosteroids Reciprocates Heavy Met als Induced Oxidative Stress in Radish by Regulating the Expression of Key Antioxidant Enzyme Genes. Braz Arch Biol Tech **61** 14. http //dx.doi.org/10.1590/1678-4324-2018160679. Shahan R 2019 The future is now: Gene expression dynamics at single cell resolution. Plant Cell **31** 933–934.

Singh I and Shono M 2005 Physiological and molecular effects of 24-epibrassinolide, a brassinosteroid on thermotolerance of tomato. *Plant growth Regul* **47** 111-119.

Song G, Hsu PY and Walley JW 2018 Assessment and refinement of sample preparation methods for deep and quantitative plant proteome profiling. *Proteomics* **18** e1800220.

Steber C M and McCourt P 2001 A role for brassinosteroids in germination in *Arabidopsis*. *Plant Physiol* **125** 763–769.

Stes E, Depuydt S, De Keyser A, Matthys C, Audenaert K and Yoneyama K 2015 Strigolactones as an auxiliary hormonal defence mechanism against leafy gall syndrome in *Arabidopsis thaliana*. *J Exp Bot* **66** 5123 - 5134. doi: 10.1093/jxb/erv309.

Sun S, Chen D, Li X, Qiao S, Shi C, Li C, Shen H and Wang X 2015 Brassinosteroid signaling regulates leaf erectness in *Oryza sativa* via the control of a specific U-type cyclin and cell proliferation. *Dev Cell* **34** 220-228.

Sun L, Yu S and Zhao F 2018 Effects of salt stress on polyamines and hormone metabolism in Grape seedlings. *Xinjiang Agric. Sci.* 55 66 - 73.

Szekeres M, Nemeth K, Koncz-Kalman Z, Mathur J. and Kauschmann A 1996

J. Indian bot. Soc. Sp. Issue Vol. 100(A) 2020:123

Brassinosteroids rescue the deficiency of CYP90, a cytochrome P450, controlling cell elongation and de-etiolation in *Arabidopsis Cell* **85** 171–82.

Takahashi T 2020 Plant Polyamines. *Plant* **9** 511 Doi:10.3390/plants9040511 www.mdpi. com/journal/plants.

Takahashi F, Suzuki T, Osakabe Y, Betsuyaku S, Kondo Y, Dohmae N, Fukuda H, Yamaguchi-Shinozaki, Kazuko S and Shinozaki Kazuo 2018. A small peptide modulates stomatal control via abscisic acid in long-distance signalling. *Nature*. **556** (7700) 235–238.

Thompson M J, Meudt W J, Mandava N B, Dutky S R, Lusby W R and Spaulding, D W 1982 Synthesis of brassinosteroids and relationship of structure to plant growthpromoting effects. *Steroids*, **39** 89–105.

Vercruyssen L, Gonzalez N, Werner T, Schmulling T and Inze D 2011 Combining Enhanced Root and ShootGrowth Reveals Cross Talk between Pathways that Control Plant Organ Size in *Arabidopsis Plant Physiol* **155** 1339–1352.

Yang J, Duan G, Li C, Han G, Zhang Y and Wang C 2019. The Crosstalks Between Jasmonic Acid and Other Plant Hormone Signaling Highlight the Involvement of Jasmonic Acid as a Core Component in Plant Response to Biotic and Abiotic Stresses. *Front Plant Sci* **10** 1349.

https://doi.org/10.3389/fpls.2019.01349.

Yu MH, Zhao ZZ and He JX 2018 Brassinosteroid signaling in Plant Microbe interaction. *Int Nat Mol Soc* **19(12)** 4091.

Yu Z, Jia D and Liu T 2019 Polyamine Oxidases play various roles in plant development and abiotic stress tolerance. Plants 8 184; doi:10.3390/plants8060184www.mdpi.com/ Poonam Meena, Sampat Nehra and P. C. Trivedi

journal/plants.

Zhang S, Cai Z and Wang X 2009 The primary signaling outputs of brassinosteroids are regulated by abscisic acid signaling. *Proc Natl Acad Sci* USA, **106** 4543–4548.

Zheng L, Gao C, Zhao C, Zhang L, Han M, An N and Xiolin R 2019 Effects of Brassinosteroid Associated with Auxin and Gibberellin on Apple Tree Growth and Gene Expression Patterns. *Hort Plt Jour* **5(3)** 93-108.

Zhu T, Tan WR, Deng XG, Zheng T, Zhang DW and Lin HH 2015 Effects of brassinosteroids on quality attributes and ethylene synthesis in postharvest tomato fruit. *Post Harvest Biol Technol*, **100** 196–204.