TOXIN PRODUCTION BY CURVULARIA LUNATA VAR. AERIA CAUSING LEAF BLIGHT IN FRENCH BEAN (PHASEOLUS VULGARIS)1

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ABSTRACT

Curvularia lunata, highly pathogenic to bean leaves produced thermostable toxic principle in synthetic media which induced symptoms similar to the disease. Toxin production was maximal after 12 days of incubation in Fries modified No. 3 medium at 30°C. Cultural conditions greatly influenced toxin production. Sucrose served as the best carbon source and ammonium sulphate and ammonium tartarate as the best nitrogen source for toxin production. The findings also confirmed that the toxin produced by Curvularia lunata is non specific.

INTRODUCTION

Visible symptoms of leaf blight of french bean (Phaseolus vulgaris) are caused not only by Curvularia lunata var. aeria but also by the toxin produced by it. The first visible symptoms could be observed 4 to 5 days after inoculation as slight chlorotic and faint brown spots. After 7 days, dark lesions had appeared and 8 days after inoculation extensive chlorosis and necrosis occurred in large areas of the leaves. This rapid rate of symptom development led us to the hypothesis that a toxin is involved in symptom development. this paper, we report our results on the optimum condtions required for production of the toxin.

MATERIAL AND METHODS

C. lunata var. aebria was isolated from diseased leaves of local fields. The fungus was grown on a shallow layer of

Fries modified 3 medium (Pringle and Scheffer, 1963) containing 30 g sucrose, 5 g ammonium tratarate, 1 g NH₄NO₃, 1 g KH₂PO₄, 0.5 g MgSO₄. 7H₂O, 0.1 g NaCl, 0.13 g CaCl₂. 2H₂O, 1 g Difco yeast extract and distilled water to 1000 ml. Aliquots of 50 ml of basal medium were dispensed in 500 ml Erlenmeyer flasks, sterilized, inoculated and kept for 16 days at 30 oC. After incubation of 2, 4, 6, 8 10, 12 14 and 16 days the mycelial mats were harvested and washed several times with distilled water so as to remove the salts and colloidal particles associated with the mycelim. Filter papers containing mycelial mats were dried at 70°C in an electric oven for 3 days to obtain constant weight and reweighed. Culture filtrate was obtained, final pH was noted and used for toxin assays after centrifugation at 5000 xg.

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Before testing for toxin content the culture medium was filter through

0.22u bacterial filter. Toxin assay was suspending host cuttings. Plants were grown in soil at room tem-Petioles were excised from perature. the stem under water and the cuttings immediately placed in serial dilutions of the toxic preparations under continuous cool white flourescent light at room temperature. The highest dilution at which necrosis of the leaf blade was observed after 48 hours was recorded as the end point; the reciprocal of this end point was taken arbitrarily as the number of activity units of the toxin per ml of the original solution e.g 1/10-10 units ml-1.

Uninoculated Fries modified 3 medium was used as control. Each value represents average of 3 replicates with 3 plants each. The influence of environmental factors (temperature, pH, carbohydrate concentration, and nitrogen concentration) on toxin production was tested in a like manner.

OBSERVATIONS

I. Time course of Toxin Production:

The results in Fig. 1 revealed that the culture filtrate of C. lunata contained toxic principle, which was first detected after 4 days at a dilution of 1/4. The toxin production rose from this value to 1000 units ml⁻¹ in 12 days and later on remained constant. Mycelial dry weight followed a similar trend but it gradually decreased after reaching a maximum in 14 days. Whereas the amount of toxin remained constant up to the end of the incubation period and the toxin titre remained stable for at least 30 days of incubation.

II. Effect of temperature on toxin production:

Toxin production was maximum at 30 °C (Fig. 2). The pathogen failed to

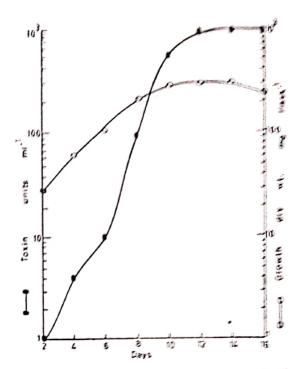


Fig. 1: Time course curve of toxin production by C. lunata

grow at and below 10°C and above 40°C temperature. The toxin in the culture filtrate was heat stable (121 °C for 15 minutes at 15 psi)...

III. Effect of pH on toxin production :

Toxin production was maximum at pH 5.0. There was a sharp decline in the toxicity of culture filtrate above and below the optimum pH (Fig. 3).

IV. Effect of sugars:

When various carbohydrates were substituted at equal sources content basis in Fries modified 3 medium, sucrose induced maximum toxin production which was followed by maltose, glucose, fructose starch, arabinose and glacatose. Raffinose was a very poor source. However, growth was maximum in arabinose followed by maltose sucrose, glucose, galactose starch, fructose and raffinose (Table I).

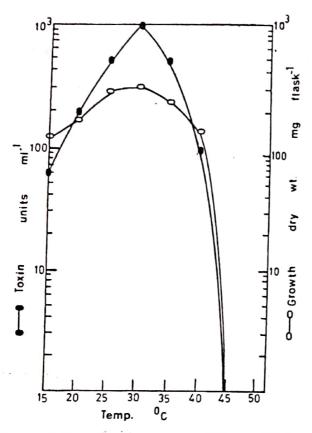


Fig. 2. Effect of temperature on toxin production and growth of G. lunata

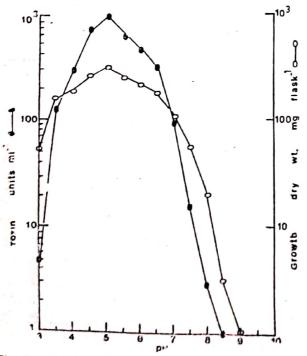


Fig. 3. Effect of pH on toxin production and of C. lunata.

TABLE I

EFFECT OF DIFFERENT CARBOHYDRATES IN INDUCTION
OF TOWN, PROPRIETORY BY CURVIN ARIA LUNATA

OF TOXIN PRODUCTION BY CURVULARIA LUNATA

(AFTER 15 DAYS OF GROWTH)

Carbon source	Growth mg flask-1	Toxin units ml-1		
Glucose	190	450		
Sucrose	320	1000		
Fructose	175	320		
Maltose	340	600		
Arabinose	365	150		
Raffinose	150	10		
Galactose	190	100		
Starch	185	200		
••				

Effect of various concentrations of sucrose on toxin production was investigated (Table II). Toxin production increased with increase in amount of sucrose from 1 to 3% but further increase in the amount of sucrose up to 10% had no effect on toxin production. However, mycelial dry weight increased

TABLE II

EFFECT OF DIFFERENT CONCENTRATIONS OF SUCROSE
ON FOXIN PRODUCTION BY CURVULARIA LUNATA

(AFTER 15 DAYS OF GROWTH)

Per cent sucrose concentration	Growth mg flask-1	Toxin units ml-1
1	155	5
1.5	22 1	. 89
. 2	290	250
2.5	310	500
3	355	1000
5	365	1000
10	410	1000

progressively in media containing upto 10% sucrose.

V. Effect of nitrogen source:

Table III revealed that among the various nitrogen sources inorganic nitrogen sources were superior to organic nitrogen sources. Ammonium sulphate, ammonium tartarate, sodium nitrate, ammonium nitrate and ammonium oxalate markedly favoured toxin production by G. lunata.

TABLE III

EFFECT OF DIFFERENT NITROGEN SOURCES ON TOXIN
PRODUCTION BY CURVULARIA LUNATA (AFTER

15 DAYS OF GROWTH)

Nitrogen sources	Growth mg flask-1	Toxin unitsml-1
Ammonium sulphate	210	1000
Ammonium nitrate	258	750
Sodium nitrate	270	840
Ammonium tartarate	365	1000
Ammonium chloride	262	500
Potassium nitrate	255	250
Asparagine	405	35
Glutamic acid	285	10
Glycine	305	20
Tryptophane	362	60

VI. Dilution end point:

Fifteen day old culture filtrate was highly active up to 1.80 dilution but further dilution reduced the activity markedly and 1:1000 was the dilution end point (Table IV).

VII. Host specificity:

Table V shows the reaction of various plant species to fungal toxins. Red

kidney bean (Phaseolus vulgaris L) was highly senstive to the toxin. Cowpea and broad bean were not infected by the pathogen but were extremely sensitive to the toxins showing total collapse and chlorosis within 48 hours. All other test plants were entirely or almost unaffected by the fungus and its toxin.

DISCUSSION

It has been observed that C. lunata produced toxic substances; the toxin production rose to maximum on the 12th day in culture medium and remained stable for at least 30 days. Luke and Wheeler (1955) observed that maximum toxin production by Helminthosporium victoriae occurred between 13 to 21 days and then declined. Smedegard and Nelson (1969) reported maximum toxin production by H. maydis after 13 to 17 days in Fries medium. Nisa and Bisen (1982) found that Coniella granati produced maximum toxin after 25 days of incubation. Bisen et al. (1983) observed that maximum toxin production by three pathotypes of Pseudomonas phaseolicola on the 9th day. Onesirosan et al. (1975) reported maximum toxin production by Corynespora cassiicola between 12 to 30 days.

Culture filtrate retained its activity even after autoclaving at 121° C for 15 minutes which indicated its thermostable nature. Thermostability appears to be a basic characteristic of toxin (Luke and Wheeler, 1955; Das and Srivastave, 1969; Sundheim, 1972 Maity and Samaddar, 1977; Vijaya Kumar and Rao, 1979).

Naef-Roth (1972) noticed that lycomarasmin one of the toxins of wilt inducing species of Fusarium, when injected to the stem of tomat plant injures the leaves severely, whereas the stem remains undamaged. Ballio et al. (1972), Main and Pero (1972) and

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TABLE IV effect on bean leaf of a culture filtrate (15 days old) of $\it CURVULARIA\ LUNATA$

Concentration of culture filtrate	Phytotoxic damage rating* at time t (hours)							
	Culture filtrate			Medium (Control)				
	24	48	72	96	24	48	72	96
Cellfree culture filtrate	5	5	5	5	0	0	1	1.5
1:1	5	5	5	5	0	0	0	0
1:10	5	5	5	5	0	0	0.5	0.5
1 : 20	4	5	5	5	0	0	0.5	1
1:40	4	5	5	5	0	0	0.5	0.5
1:80	3	4	5	5	0	0	0	1
1 : 160	2	3	4	-5	0	0	0	.0
1 : 320	2	2.5	3	5	0	0	0.5	1
1:640	1.5	2	3	4	0	0	0.5	0.5
1 : 750	1	1.5	2	3	0	0	0	0
1:1000	0.5	1	1.5	2	0	0	0	0

Cutting from 2 week old seedlings

TABLE V

RELATIONSHIP BETWEEN SUSCEPTIBILITY TO THE CULTURE FILTRAGE OF CURVULARIA LUNATA AND DISEASE REACTION OF VARIOUS PLANTS (15 DAYS OLD CULTURE FILTRAGE)

Plant species		*Phytotoxic damage rating				
	Culture	filtrate	Medium (control)			
	24 hours	48 hours	24 hours	48 hours		
Red Kidney	3	4.5	0	0.5		
Cowpea	2.5	4	0.5	1		
Broad bean	1	1.5	0	0		
Tomato	0	0.5	0	0.5		
Brinjal	0	1	0	0.5		
Chillies	0	1	0	1		
Pca	0	0	0	0		

*Cuttings from 2-3 week old seedlings

^{*} Rating scale 0=no symptoms to 5=complete chlorosis and collapse of plant cuttings were rated after exposure for indicated time. Each value represents the average of 3 replicates with 3 cuttings each.

^{*} Rating scale 0—no symptoms to 5— complete chlorosis and collapse of cuttings. Cuttings were rated after 24 and 48 hours of exposure.

Each value represents average of 3 replicates with 3 plants each.

Schlosser (1972) reported that the host tissues showed disease symptoms when the tissues were infected by the toxins produced by the pathogen. Bean leaves were affected by cell free culture filtrate obtained from .15 day old culture.

Our data further confirmed that the toxin produced by G. lunata is a nonsspecific one as it expressed symptoms on various other plants.

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