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ANTIMYCOTIC ACTIVITIES OF SOME RHIZOSPHERE FUNGI AGAINST *MYROTHECIUM RORIDUM* CAUSING LEAF SPOT OF COTTON

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A study was undertaken to investigate the antimycotic activities of some rhizosphere fungi of cotton against *Myrothecium roridum*. A total of five species viz. *Aspergillus flavus*, *A.niger*, *Trichoderma viride*, *T.harzianum* and *P.oxalicum* were tested for antimycotic activity against *Myrothecium roridum* by dual culture method in *in vitro*. There was little difference among the rhizosphere fungi in causing inhibition of the pathogen after three days of inoculation in dual culture, but significant difference was observed on 5th days of incubation. Fast growing antagonist was found to be most promising than slow growing antagonist. Among the five antagonists tested *in vitro* against *Myrothecium roridum*, *T.harzianum* was found to be most effective as compared to others.

Key words: Antimycotic, cotton, Myrothecium, rhizosphere, Trichoderma, Penicillium.

The constant use of fungitoxic chemicals adds to the environmental pollution, consequently, efforts are under way for finding alternatives to chemical fungicides. Microorganisms which establish in the rhizosphere are ideal candidates for use as biocontrol agent, since rhizosphere provides the front line defense for root against attack by pathogens (Montensions *et al.*2002).The control of plant disease by chemicals has been subject of public concern and security due to the possible harmful effect on the environment and their undesirable effect on the nontarget microorganisms (Gerlagh *et al.* 1999).

Biological management is the control of microorganisms by another, which reduces the inoculums density of disease producing activities of pathogen by one or more organisms. Application of biological agents for reduction of diseases is an integral component of cultivation. Weindling (1932) started the first work on *Trichoderma* and discovered that *T.lignorum* controls the growth of number of soil borne fungi. *Trichoderma* and their strains or isolates are known to differ in their potential to control the disease (Venkat Kota, 2006).*Trichoderma* are ubiquitous, soil borne,greenspored ascomycetes with multiple attributes in biocontrol,enzyme production and novel antibiotics (Siameto *et al.* 2010).

Biocontrol is one of the advanced technique utilized in agricultural field, which is found to be more selective and with no side effects. It is cheap, Biological agents are self propagating and self perpetuating there is no risk of resistance and are non-hazardous. The biocontrol fungi have a potential to restrict other fungi and arrest of pathogen achieved by way of antibiosis, mycoparasitism and competition.

The main aim of the present study was to evaluate the antimycotic properties of some rhizosphere fungi against *Myrothecium roridum* causing leaf spot cotton.

MATERIALS AND METHODS

a) Isolation of Pathogen :

Infected leaves of cotton were collected from various cotton fields in Amravati region. *Myrothecium roridum* was isolated from diseased leaves of various cotton varieties and maintained on (Potato Dextros Agar) PDA slants. PDA have been selected to maintain pure cultures of isolates in the research laboratory. The isolates viz. *Myrothecium roridum, Aspergillus flavus ,A.niger,Trichoderma harzianum,T.viride and Penicillium oxalicum* were identified from available literature. Pathogenecity test was confirmed in the laboratory on potted cotton plants by Koch's postulate method.

b) Isolation of Antagonists :

Antagonistic fungi were collected from the rhizosphere of cotton plants during the rainy season by serial dilution method (Waksman,1927). Only five fungi were selected for antagonistic study viz. *Asper*-

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gillus flavus, A.niger, Trichoderma harzianum, T.viride and Penicillium oxalicum.

c) Antagonistic Study :

The dual culture method was adopted to observe the antimycotic effect of different isolates of the test antagonists. Test antagonists were screened against the pathogen Myrothecium roridum in dual culture on potato dextrose agar medium in petriplates. Autoclaved PDA medium was poured into petriplates and allowed to solidify. 5 mm disc of pathogen was kept in centre of petriplate while antagonists were inoculated 4 cm apart from it on PDA. In control only disc of the pathogen was inoculated. The petriplates were inoculated at 27.2°C. The radial mycelial growth of colony in diameter were measured on 3rd,5th and 7th days after incubation. Mycelial diameter of each treatment was compared with control plates. The percent inhibition of mycelial growth was calculated by using formula-



Where, TFC= Test fungus in control TFTr= Test fungus in treatment.

RESULTS AND DISCUSSION

The data from Table 1 showed that *Trichoderma harzianum* was more effective which control the radial mycelial growth of the pathogen Myrothecium roridum upto 58.33 percent on 3rd day of incubation followed by Trichoderma viride (38.88%). Aspergillus flavus and Penicillium oxalicum showed less than 31 percent growth inhibition. Aspergillus niger was found to be least effective (16.16%) on 3rd day of incubation. Trichoderma harzianum showed 100 per cent growth inhibition on 5th day of incubation followed by Aspergillus flavus(97.22%), Penicillium oxalicum(95.45%), Trichoderma viride(93.18%) and Aspergillus niger (84.09%). But as the days progressed in incubation, all the five antagonistic fungi showed 100 per cent growth inhibition upto 7th day of incubation. The radial mycelia growth of pathogenic fungi Myrothecium roridum was inhibited more effectively since 3rd day of incubation by Trichoderma harzianum.

From the present investigation it is clear that *Trichoderma harzianum* showed maximum growth inhibition right from the beginning as compared to other four tested antagonistic fungi. Upadhyay and Mukhopadhyay (1986) noted that *Trichoderma harzianum* produces extracellular enzymes $\beta(5,3)$ gluconase and chitinase which are capable of degrading cell walls of pathogenic fungi. Sonawane and Pawar (2001) reported that *Trichoderma harzianum* was highly effective in controlling the vegetative growth of the pathogen.

Arumugam *et al.*(2003) studied the antagonistic effect of *Trichoderma harzianum*, and isolated the enzyme

Sr.	Antagonistic fungi	Radial mycelial growth (mm)*			% growth inhibition		
N0		3 DAI	5 DAI	7 DAI	3 DAI	5 DAI	7 DAI
1	Aspergillus flavus	25.00	1.00	0.00	30.55	97.72	100.00
2	Aspergillus niger	30.00	7.00	0.00	16.66	84.09	100.00
3	Trichoderma viride	22.00	3.00	0.00	38.88	93.18	100.00
4	Trichoderma harzianum	15.00	0.00	0.00	58.33	100.00	100.00
5	Penicillium oxalicum	25.00	2.00	0.00	30.55	95.45	100.00
6	Control	36.00	44.00	51.00	-	-	-

Table 1 : Effect of different antagonists on radial mycelial growth of Myrothecium roridum.

*Mean of three replicates. DAI - Days After Incubation.

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chitosanase. They pointed that the enzyme chitosanase inhibited the conidial germination of *Myrothecium verrucaria*. Therefore it is clear that enzyme secreted by the antagonist shows potential antimycotic property which inhibits the growth of pathogen. Similar findings have been observed by Brozova (2004) and Gupta (2004).

Chindambaram *et al.*(2004) also found that bioagents like *Trichoderma* were significantly effective for controlling cotton diseases.

The present study indicates that management of diseases by potential biocontrol agent like *Trichoderma harzianum* is effective against *Myrothecium roridum*. It is eco-friendly, non-hazardous and safe to environment.

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REFERENCES

Arumugam P, Pallavi P and Balsubramanian R 2003 Production and properties of extra cellular chitosanase lysozyme of *Trichoderma harzianum.J Mycol Pl Pathol* **33** (3) 414-420.

Brozova J 2004 Mycoparasite fungi *Trichoderma sp.* in plant protection. *Plant Protection Sources*.**40**(2)63-74.

Chindambaram P, Johnson I, Kannan A and Babu S 2004 Biological control of cotton fungal diseases using formulations of *Trichoderma* and *Pseudomonas*. International Symposium on "*Strategies for Sustainable Cotton Production- A Global Vison*". 23-25 Nov. 2004. University of Agricultural Sciences. Dharwad. *Crop Protection* (**3**) 344-47.

Gerlagh MK ,Goosan-vande K, Geijn HM, Fokkemma NJ, and Vereijken PFJ 1999 Long term biosanitation by application of *Coniothyrium minitans* on *Sclerotinia sclerotiorum* infected crops. *Phytopathology*.**89** 141-147.

Gupta V 2004 Compatibility of biocontrol agents *T. harzianum* with pesticides. *J Mycol Pl Pathol* **34**(2) 132-134.

Montensions E, Bonaterra A, Bodasa E, Frances J, Alemany J, Lioernte I and Moragrega C 2002 Plant microbe interaction and new biotechnological methods of plant disease control. *Indus Microbiol.* **5** 169-175.

Siameto E N,Okota S Amugune O N and Chege C N 2010 Antagonism of *Trichoderma harzianum* isolates on soil borne plant pathogenic fungi from Embu District, Kenya. *J Yeast Fungal Research*.**1**(**3**) 47-54.

Sonawane S S and Pawar N B 2001 Studies on biological management of chickpea wilt. *J Maharashtra Agric Univ* **26**(2)215-216.

Upadhyay J P and Mukhopadhyay A N 1986 Biological control of *Sclerotium rolfsii* by *Trichoderma harzianum* in sugar beet. *Tropical Pest Management* **32** (3)215-220.

Waksman S A 1927 *Principles of Soil Microbiology*. Williams and Wilkinson Company, Baltimore.

Weindling R 1932 *Trichoderma lignorum* as a parasite of other fungi. *Phytopathology* **22**(1)837-845.

Venkat Ramesh Kota, Kulkarni S and Yashoda R. Hegde 2006 Biological management of crown rot of Banana. *J Pl Dis Sci*, **1**(2):143-145.