

RESEARCH ARTICLE

Leaf architecture in Actinodaphne and Cryptocarya species

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Abstract: Anatomical studies and leaf architecture are the important tools for the study of plants. They also help in the correct identification of genera and species which are morphologically difficult to delimit. Identification of members of the family Lauraceae are also such morphologically challenging genera and species but of economic value. Hence, the present study was undertaken. The leaves were cleared and stained with saffranine to study the venation pattern and unique feature to find out anatomical marker by using a microscope. Four species of *Actinodaphne* and three species of *Cryptocarya* were collected and their venation pattern was studied. *Cryptocarya* is valued for its timber and other medicinal uses, *Actinodaphne hookeri* Meissn. is one such medicinal plant whose leaves have been traditionally used for diabetes and disorders of the urinary tract. Based on the leaf architecture, the species could be separated. A key to separate the species was also provided herewith. Three types of venation patterns were found namely Pinnate camptodromous Eucamptodromous, festooned brochidodromous and Acrodromous type. The highest vein orders were 5° and 7°.

Keywords: Venation, Pinnate Camptodromous, Eucamptodromous, festooned brochidodromous, Acrodromous.

Introduction

The system of descriptive terminology and leaf architecture have been presented by Madler and Strauss (1971) Ferguson (1971) Walther (1972) and others Krusmann (1960), Stace (1965) and Mouten (1960, 1967). An authentic terminology of leaf architecture has been given by Hickey (1973, 1979). Dilcher (1974) presented the outline of the leaf architecture of dicot leaves. Hickey and Wolfe (1975) and Melville (1976) have revised the data and classified the architecture of leaves of dicotyledonous plants. Hall and Melville (1951) proposed a veinlet termination number as a technique for testing the purity of fragments of a particular leaf type for pharmacognostical properties. Thus, anatomy and leaf architecture study can be used as an additional tool to help in the correct identification of a species. The morphological characters of Lauraceae are well explained but key to members of the Lauraceae and Cryptocarya is based on the development of anther

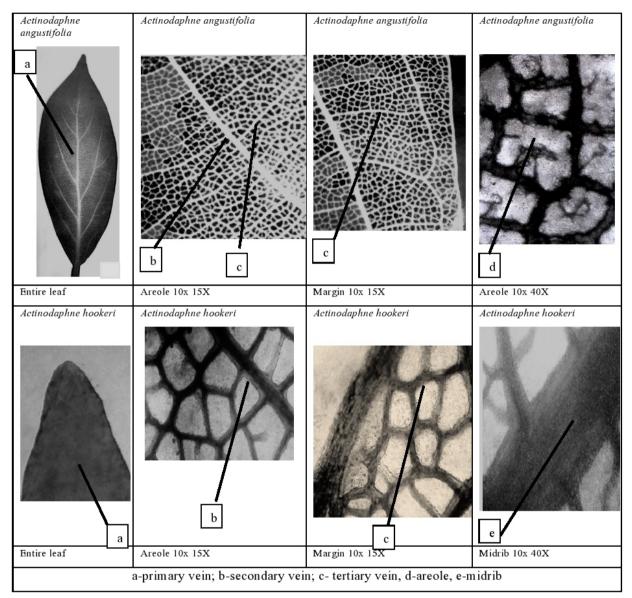
Meenakshi Vaidya meenakshi.vaidya@mithibai.ac.in and stamens, which makes the correct identification difficult (Hooker 1883) and hence the present study was undertaken.

Material and methods

The collection of plant material for *Cryptocarya* was done from the species growing naturally at Shillong- Meghalaya. The collected plant material was identified and checked with the help of Standard Herbaria from A.R.I. Herbarium, Pune and B.S.I. Eastern Circle, Shillong.

The method described by Payne, (1969) and Mohan Ram and Nayyar (1978) in which both fresh as well as dried material was used for the study of leaf architecture. The material was dried between blotting papers and then used. The leaves fresh or dried were first cleared by keeping them in 5% sodium hydroxide solution at room temperature for 1-2 days. The decolored leaves were washed and transferred to 5% sodium hypochlorite till they were transparent. The leaves were then thoroughly washed to remove traces of hypochlorite and stained with aqueous saffranine by keeping them in it for 10-15 minutes and mounted in glycerine jelly. For anatomical studies, the terminology used as

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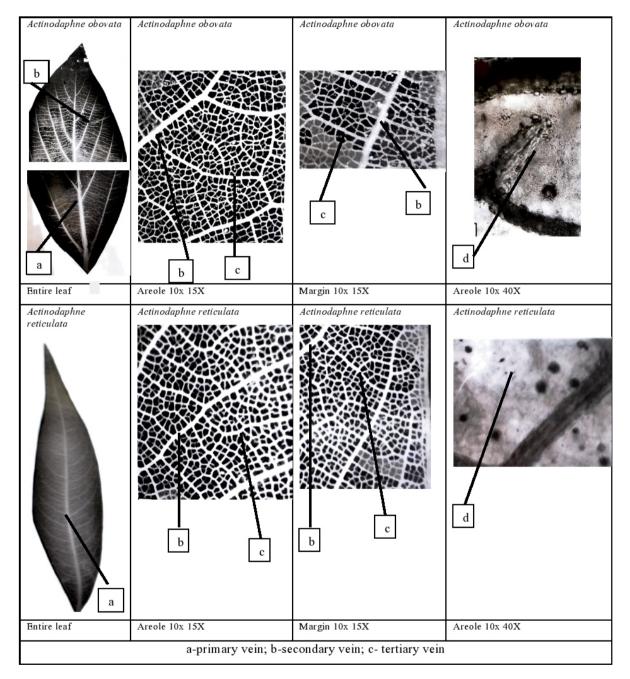


suggested by Hickey (1973, 1979), Hickey and Wolfe (1975), Dilcher (1974) and Melville (1976).

Observations

Four species of *Actinodaphne* are studied namely *Actinodaphne angustifolia*, *A. hookeri*, *A. obovata* and *A. reticulata*. Three species show Pinnate Camptodromous eucamptodromous secondaries. The highest vein order is 5°. The angle of divergence of secondary veins is acute moderate in *A. reticulata*, while it is acute narrow in *A. obovata* and *A. angustifolia*. Further, these species have random reticulate pattern of tertiary veins. The angle of origin of tertiary veins seen is RR/AR/AO/AA in *A. reticulata*, and tracheoids are absent. Pattern is percurrent in *A. obovata* and *A. angustifolia*. These are separated on the basis of angle of origin of tertiary veins which is RR/AR/RA/OR in *A. obovata* and AA/AR/RR/RA/OA in *A. angustifolia* with tracheoids present in both. Thus, the three species are separated from each other on the basis of the leaf architecture patten in the genus *Actinodaphne*. *A. hookeri* has Acrodromous venation. The highest vein order is 5°. The angle of divergence of secondary veins is acute narrow. The angle of origin of tertiary veins seen is RR/RO/RA in *A. hookeri*.

Key to separation of species of Actinodaphne:



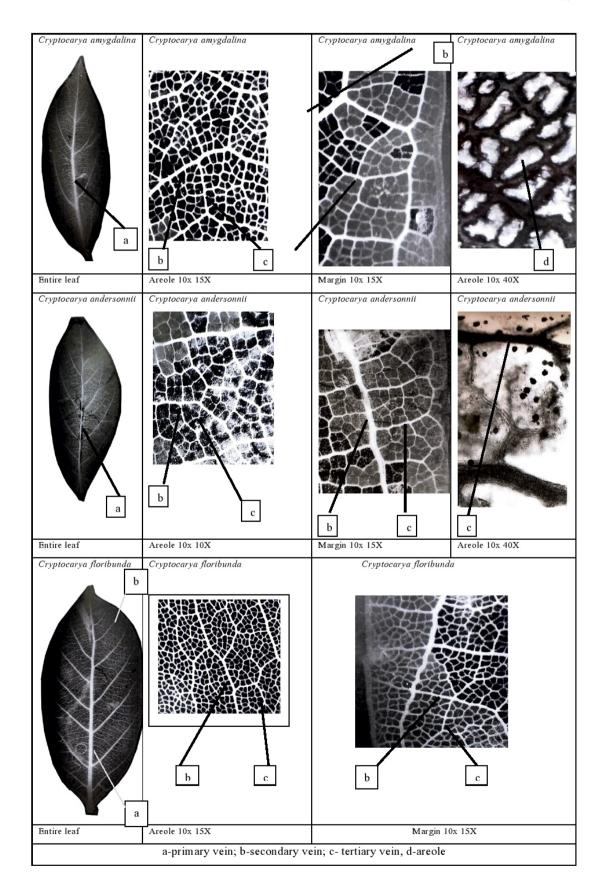
Pinnate Camptodromous

Eucamptodromous secondaries

Secondary veins angle of divergence acute narrow Highest vein order 5° Pattern of tertiary veins percurrent

Tracheoids absent Angle of origin of tertiary veins AA/AR/RR/RA/OA ----- A. angustifolia Angle of origin of tertiary veins RR/AR/AO/AA------A. reticulata Tracheoids present Angle of origin of tertiary veins RR/AR/RA/OR ------ A. obovata

Acrodromous Secondary veins angle of divergence acute



narrow

Highest vein order 5°
Pattern of tertiary veins percurrent
Tracheoids absent
Angle of origin of tertiary veins
RR/RO/RA A.
hookeri

Three species of Cryptocarva are studied namely Cryptocarya andersonni, C. floribunda and C. amygdalina. The type of venation is pinnate Camptodromous with festooned brochidodromous secondaries. Secondary veins angle of divergence acute narrow and pattern of tertiary veins percurrent. Tracheoids are absent in all three species. However, these species can be separated from each other on the basis of angle of origin of tertiary veins RR/OR/RA/OO/AA/OA in C. andersonni and AR/AA/RR/RA in C. floribunda and the highest vein order is 5° in both. C. amygdalina shows RR/RO/OR/OO/RA/OA with an additional character of the highest vein order being 7°. [A-Acute angle, R-Right angle, O-Obtuse Angle]

The three species of *Cryptocarya* can be separated on the basis of venation and the key to their separation is:

Pinnate Camptodromous

Festooned brochidodromous secondaries Secondary veins angle of divergence acute narrow

Highest vein order 5° Pattern of tertiary veins percurrent Angle of origin of tertiary veins RR/OR/RA/OO/AA/OA------ *C. andersonni* Angle of origin of tertiary veins AR/AA/RR/RA------ *C. floribunda*

Highest vein order 7° Pattern of tertiary veins percurrent Angle of origin of tertiary veins RR/RO/OR/OO/RA/OA------ *C. amygdalina*

References

Chodankar and Vaidya 2021 a: Study Of Leaf Architecture In Eleven Species Of *Cinnamomum* Blume. Of Family Lauraceae In WJPR Volume 10, Issue 2, 2021: 1190-1199. ISSN: 2277-7105

Chodankar and Vaidya 2021 b Patterns Of Leaf

Architecture In Six Species Of *Phoebe* From Family Lauraceae In WJPR **10 (3)** 1772-1778

Chodankar and Vaidya 2021 c Anatomy – A Useful Tool In The Identification Of Ten Species Of Genus *Machilus* Of Family Lauraceae. *J. Indian bot. Soc.* **101** (4) 349-355

Dilcher D L 1974 The Study of Angiosperm Leaf remains. *The Botanical Rev.* **40** 1-157.

Ettingshausen C 1861 Die Blattskelete der Dicotyledonen mit besonderer Ruckicht auf die untersuchung and Bestimming der fossilen Pflanzengeste. Kon. Huf. and Staatsdruckeri Wein: 308.

Ferguson D K 1974 On the taxonomy of recent and fossil species of *Laurus* (Lauraceae). *J. Link.Soc. Lond. Bot.* **68** 51-72.

Hall JP and Melville C 1951 Veinlet termination number a new character for the differentiation of leaves. J. Pharm. Pharmac., **3** 934-941.

Hickey L J 1973 Classification of the architecture of dicotyledonous leaves. *Amer. J. Bot.* **60** 17-35.

Hickey L J 1979 A revised classification of the architecture of dicotyledonous leaves. In: *Metcalfe and Chalk.* Anatomy of dicotyledons. Clarendon Press, Oxford.

Hickey L J and J A Wolfe 1975 The basis of angiosperm Phylogeny: Venation. *Ann. Mlssouri Bot.Gard.*,**62** 538-589.

Hooker J D 1883 Flora of British India Vol-5 116-189. Reeve and Co., London.

Krusman G 1960 Handbuch der Laubge Holze I and II. Berlin.

Madler K and Strauss A 1971 Em system der blattformen mit spezieller anwendung für die bestimmung Noegener blattreste. *Bot. J.* **90** 562 - 574.

Melville R 1976 The terminology of leaf architecture of Apocynaceae. Taxon, **25** 549-561.

Mohan Ram H Y and Nayyar Vijaylaxmi 1978 Leaf clearing technique with a wide range of applications; Proc. Indian Acad. Sci. (Plant Sci.) B. **87** 125-127.

Mouten J A 1960 Sur la systeatique foliare en paleobotanique. Bull. de la Ser. *Bot. fr.* **113** 492 - 502.

Mouten J A 1967 Architecture de la nervation Foliare.

92. Congress national des societetes savantes. Strashourg et Columr - III: 165 - 176.

Payne W W 1969 A quick method for clearing leaves. Ward's Bulletin **8(61)** 4,5.

Stace C A 1965 Cuticular studies as an aid to plant taxonomy. *Bull. Bri. Musi. Bot.*, **4(1)** 1-78.

Vaidya M 2014 Anatomy- Leaf Venation: A tool for identification of some species of genus *Litsaea* Lamk. of family Lauraceae. *J. Indian Bot. Soc.*, **93** (**3 & 4**) 103-108

Vaidya M 2015a Role of Anatomy in identification of twelve species of genus *Litsaea* Lamk. in *Asian Journal of Biochemical and Pharmaceutical Research*, **5(1)** 93-100.

Vaidya M 2015b Study of leaf architecture in some species of *Litsaea* Lamk. Of family Lauraceae in Journal of anatomy. *Photon* **115** 182-184.

Vaidya M 2016 b Study of Trichomes in Some Species of *Litsaea* Lamk. in WJPR, **5 (2)** 1069-1077.

Vaidya M 2016a Study of Stomatal Complexes in Some Species of the Genus *Litsaea* Lamk. in WJPR, **5(1)** 851-863.

Walther H 1972 Studien uber tertiare Acer Milteleuro Pas. Abh. Staaatl. Mus. Mineral. Geol. Dresdan, **19** 1 -309