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INDUCED COMB-COLOUR MUTANTS IN COCKSCOMB (CELOSIA CRISTATA L.)

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8 Abstract We have isolated twelve comb-colour mutants in cockscomb induced through 9 conventional as-well-as in vitro mutagenesis. The spectrum of these mutants included the combs with rose-red, orange, vivid red, coral red, vermillion and carmine colour. These comb-10 11 colours were obviously distinct from that in control (magenta). The content of total betalain, amaranthin, betanin, betalamic acid and betaxanthin was reduced in the mutants. Notably, the 12 relative contents of these pigments in the mutant also differed, which might be the reason for 13 variation in comb-colour of these mutants. Therefore, the mutants isolated in the present 14 investigation can, used for crop improvement and serve as a model to study betalain 15 16 biosynthesis in cockscomb.

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18 Keywords: EMS, gamma ray, in vitro mutagenesis, mutagen, ornamental

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20 Running title: Colour mutants in cockscomb

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23 Introduction

24 Celosia cristata (L.) or cockscomb is widely planted in the gardens of South America, Africa, 25 and Asia (Bakar et al. 2014). The compact form and velvety texture of its inflorescence makes cockscomb a plant suitable for landscaping and roadside plantation (Meng et al. 2009). 26 27 Cockscomb is the focal point of a garden due to the unique shape and fascinating hues of its inflorescence due to the presence of betalain pigments, which are restricted to the few members 28 29 of Order Caryophyllales (Warhade and Badere 2018). Betalains have at least one heterocyclic nitrogen atom in their ring. However, they are not alkaloids sensu stricto as they are acidic in 30 nature due to the presence of several carboxyl groups. Conjugated double bonds in the structure 31 of betalains impart colouring properties to the molecules. Based on the colour, two major 32 classes of betalains are identified viz., purple coloured betacyanins and yellow to orange 33 34 coloured betaxanthins (Harris et al. 2012). The combination of these betacyanins and 35 betaxanthins generates an array of hues in the plant part synthesising the betalains.

Mutation breeding is an effective method of crop improvement. It modifies one or two desirable traits, like flower colour, of the already established variety without altering the other traits (Ahloowalia et al. 2004). Coupling of induced mutagenesis with in vitro techniques has been suggested to increase the frequency of mutations. The technique of micropropagation supplements induced mutagenesis by amplifying the mutated sector thereby increasing the frequency of mutation (Maluszynski et al. 2001; Nasri et al. 2021).

42 We initiated a study to compare the effectiveness and efficiency of conventional mutagenesis 43 vis-à-vis in vitro mutagenesis in cockscomb (Rinkey and Badere 2023). In this study, we treated the seed of cockscomb with either EMS or GR to raise the M₁ (conventional 44 mutagenesis) and RM₁ (in vitro mutagenesis) population. This population was harvested on 45 46 maturity and the seeds thus obtained were sown in next season to raise the M₂ and RM₂ generation. The M₂ and RM₂ populations were thoroughly screened to isolate the putative 47 mutants for various traits. The seeds of these putative mutants were sown to raise the M_3 and 48 RM₃ generation to study their inheritance thereby confirming the mutant. Among others, a few 49 comb-colour mutants in cockscomb were isolated in this investigation. The present paper 50 describes the morphological and phytochemical characterisation of the comb-colour mutants 51 52 isolated during the investigation.

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54 Materials and Methods

55 Isolation of comb colour mutants

Two modes of mutation induction viz. conventional and in vitro were employed in the present 56 57 investigation. For this, the seeds of cockscomb treated with either EMS or gamma ray as described earlier (Rinkey and Badere 2023, 2024). Subsequently, the M₁ and RM₁ generations 58 were raised and harvested plant-wise. In the subsequent season, the seeds harvested from M₁ 59 and RM₁ plants were sown in a tray along with control to raise the M₂ and RM₂ generations on 60 plant-to-row basis. Enough seedlings at 4 leaf stage were transplanted in the experimental plots 61 to maintain 25 plants per concentration/dose. The population was thoroughly screened to 62 isolate the putative mutants for various traits. In the next season the M₃ and RM₃ generations 63 were also raised on plant-to-row basis from the seeds of putative mutants harvested in M₂/RM₂ 64 generation. The inheritance of comb-colour, for which the putative mutants were isolated, was 65 tested in this generation. Fifty seeds from each putative mutants were sown in the tray along 66 67 with the control. Subsequently, the 30d-old seedlings were transplanted to the field and allowed 68 to grow as earlier. The population was screened regularly to monitor the growth and development of plants. The putative mutant whose progenies exhibited the character for which 69 it was isolated in the M₂/RM₂ generation was isolated as viable mutant. Such viable mutants 70 71 were characterised for their morphological features.

72 Pigment analysis in the comb-colour mutants

The comb-colour mutants isolated in the present investigation were qualitatively and quantitatively analysed for their betalain pigments. The qualitative analysis was performed using thin layer chromatography, while the amaranthin, betanin, betalamic acid and betaxanthins were quantified spectrophotometrically as described by Warhade and Badere (2015). Later, the pigment contribution and pigment ratio were calculated according to the formulae given by Warhade and Badere (2015, 2018).

79 Statistical analysis

The data obtained during the present investigation were statistically analysed by computing the
arithmetic mean, standard error and Student's t value using MS-Excel.

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83 **Results**

84 Viable comb-colour mutants

The putative comb-colour mutants were induced with higher frequency in RM₂ population in response to the EMS treatment. In the same manner the frequency of putative comb-colour mutants, in response to GR treatment, was also higher in RM₂ population compared the M₂ population. Apart from this the pre-soaking of seeds prior to EMS treatment also reduced the frequency of putative comb-colour mutants (Rinkey and Badere 2023).

Seeds of putative comb-colour mutants were sown to raise the M₃/RM₃ generation. Eight plants 90 91 from M_3 families and four plants from RM_3 families displayed the character for which they were selected. These twelve plants were designated as viable mutants and characterised. All 92 but one viable comb-colour mutant was induced by EMS in M3 generation. Moreover, most of 93 the comb-colour mutants induced by EMS, in fact, were multiple mutants. The mutant 94 95 E/0/0.15/47/8/5 with rose red colour had smaller leaves and larger comb. Contrastingly another rose red mutant, E/0/0.15/47/10/8, had marginally larger leaves and a greater number of 96 97 branches compared to the control. The orange colour mutant, E/0/0.15/54/7/15, had slightly 98 larger comb. Similarly, one of the coral red mutants, E/3/0.15/30/3/10, had shorter internode & inflorescence, and a vermillion mutant E/6/0.20/19/5/1 had comb with markedly longer 99 diameter. In addition to these, EMS also induced vivid red mutant E/0/0.25/60/10/25, which 100 was similar to control for other characters studied (Table 1). Among the comb colour mutants 101 induced in RM₃ generation, R/E/0/15/2/2/15 (vivid red) was similar to the control. However, 102 103 the other three mutants were multiple mutants. While comb of EMS-induced R/E/0/20/1/1/12had carmine colour and smaller internodes, another one with vermillion coloured comb 104 (R/E/0/25/11/1/8) had longer comb diameter. Similarly, gamma ray-induced R/G/30/4/3/14 105 was dwarf with carmine comb-colour (Table 1). 106

107 Phytochemical characterisation of the comb colour mutants

108 The TLC profile of betalain pigments in control and the comb-colour showed five bands having 109 R_f values of 0.11 (magenta), 0.17 (magenta), 0.27 (reddish pink), 0.36 (light pink) and 0.49 110 (yellow). Notably, the profile of all the comb-colour mutants was alike control showing five 111 bands of same R_f and colour. However, the intensity of bands with R_f value 0.11 and 0.17 112 increased in all the mutants in comparison to control except for orange, rose red and vivid red 113 coloured mutants. Rather, the intensity of the band with R_f value 0.11 was decreased in the 114 orange mutant (E/0/0.15/54/7/15) compared to that of control (Table 2).

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Generation	Mutant	Plant height (cm)	No. of branches	Length of internode (cm)	Diameter of stem (cm)	Leaf area (cm ²)	Length of comb (cm)	Diameter of comb (cm)	Circumfe- rence of comb (cm)	Length of fertile region (cm)	Colour of comb
	Control*	29.0-80.0 (57.2)	7-16 (11)	1.7-6.9 (3.4)	0.34 - 1.75 (1.00)	15.8-36.2 (25.7)	8.2-19.5 (13.8)	1.0-8.8 (6.1)	7.5-14.0 (10.9)	6.5-14.0 (9.4)	Magenta
	E/0/0.15/47/8/5	75.0	13	4.2	0.92	10.8	13.8	11.4	17.9	7.0	Rose-red
	E/0/0.15/47/10/8	74.5	19	3.6	0.97	38.9	19.2	6.8	12.7	14.3	Rose-red
	E/0/0.15/54/7/15	70.0	8	4.0	0.90	24.5	16.0	6.9	15.0	11.0	Orange
M ₃	E/0/0.25/60/10/25	29.5	13	3.0	0.90	22.8	11.0	6.6	10.5	6.8	Vivid red
	E/3/0.15/30/3/10	65.5	15	1.5	0.53	17.8	8.0	6.5	12.0	10.0	Coral red
	E/3/0.25/23/1/5	44.0	14	2.5	0.55	16.3	10.0	5.5	8.7	7.0	Coral red
	E/6/0.20/19/5/1	76.0	11	6.0	1.08	22.1	12.0	10.0	14.0	8.0	Vermilion
	G/30/11/2/1	42.7	9	2.1	0.62	18.9	17.2	6.1	12.2	13.7	Vermilion
	Control*	23.0-80.0 (56.5)	7-16 (11)	1.7-6.9 (3.6)	0.34 - 1.25 (1.00)	10.8-36.2 (24.6)	7.2-19.5 (13.3)	1.8-8.0 (6.1)	2.5-14 (8.7)	6.5-14 (9.4)	Magenta
RM ₃	R/E/0/15/2/2/15	60.2	10	5.2	0.72	13.7	12.7	7.2	12.2	7.1	Vivid red
	R/E/0/20/1/1/12	39.1	9	1.1	0.95	11.0	8.2	3.2	9.1	7.5	Carmine
	R/G/30/4/3/14	22.7	11	3.9	1.18	12.0	14.1	2.4	5.1	9.5	Carmine
	R/E/0/25/11/1/8	46.5	8	5.1	1.01	15.2	9.5	9.1	12.5	6.7	Vermilion

Table 1. Morphological traits of the comb colour mutants

* The figures are the minimum and maximum value of the trait and the figure in parentheses is the mean value of the trait.

Subsequently, the content of betalain pigments viz. amaranthin, betanin, betalamic acid and betaxanthin was spectrophotometrically quantified in control and the mutants. The total betalain content in the mutants was either unaffected or reduced. Remarkably, the total betalain content in the mutants isolated from M_3 population was reduced except in the mutant E/3/0.15/30/3/10 with coral red colour. In contrast, the total betalain content was unaffected in the mutants isolated from RM_3 population with an exception of vermillion coloured mutant RE/0.25/11/1/8 where it was decreased (Table 3). The content of amaranthin and betanin was

Table 2. TLC profile of betalain pigments in the comb of comb-colour mutants										
		R _f value and colour of the band								
Mutant	Comb	0.11	0.17	0.27	0.36	0.49				
	colour	Magenta	Magenta	Reddish pink	Light pink	Yellow				
Control	Magenta	+	+	+	+	+				
M ₃ generation										
E/0/0.15/47/8/5	Rose-red	+	+	+	+	+				
E/0/0.15/47/10/8	Rose-red	+	+	+	+	+				
E/0/0.15/54/7/15	Orange	<	+	+	+	+				
E/0/0.25/60/10/25	Vivid red	+	+	+	+	+				
E/3/0.15/30/3/10	Coral red	++	++	+	+	+				
E/3/0.25/23/1/5	Coral red	++	++	+	+	+				
E/6/0.20/19/5/1	Vermilion	+++	++	+	+	+				
G/30/11/2/1	Vermilion	+++	++	+	+	+				
RM ₃ generation										
RE/0.15/2/2/15	Vivid red	+	+	+	+	+				
RE/0.2/1/1/12	Carmine	++	++	+	+	+				
RG/30/4/3/14	Carmine	++	++	+	+	+				
RE/0.25/11/1/8	Vermilion	+++	++	+	+	+				

reduced in almost all the mutants compared to control. The amaranthin content was reduced, in comparison to control, in all the mutants induced through conventional mutagenesis. Similarly, all the mutants induced through in vitro mutagenesis had amaranthin content lower than control except the vivid red mutant RE/0.15/2/2/15 (Table 3). In the same way all the mutants induced through conventional mutagenesis, except rose red E/0/0.15/47/8/5 and coral red E/3/0.15/30/3/10, had lower betanin content compared to the control. In contrast, the betanin content was reduced only in vermillion mutant RE/0.25/11/1/8 among the mutants

induced by through vitro mutagenesis. A marked difference in the content of betalamic acid and betaxanthin was observed between the mutants induced through conventional and in vitro mutagenesis. Most of the mutants induced through conventional mutagenesis had content of betalamic acid and betaxanthin similar to control. Exceptionally the orange mutant

E/0/0.15/54/7/15 had reduced content of both these pigments. Similarly, betalamic acid and betaxanthin content were reduced in rose red mutant E/0/0.15/47/8/5 and vivid red mutant E/0/0.25/60/10/25, respectively. While the content of betaxanthins were reduced in all the mutants induced through in vitro mutagenesis; that of betalamic acid was increased in all but except vermillion mutant RE/0.25/11/1/8 (Table 3).

Table 3. Betalain content (mg/g FW) in the comb of comb-colour mutants										
Mutant	Comb colour	Amaranthin	Betanin	Betalamic acid	Betaxanthin	Total Betalain content				
Control	Magenta	1.07 ± 0.05	0.76±0.14	0.27±0.03	0.0003 ± 0.00	2.10±0.21				
M ₃ generation										
E/0/0.15/47/8/5	Rose-red	0.86±0.05*	0.59±0.03	0.18±0.01*	0.0002 ± 0.00	1.50±0.04*				
E/0/0.15/47/10/8	Rose-red	$0.52 \pm 0.09*$	0.33±0.02*	0.24 ± 0.02	0.0002 ± 0.00	0.93±0.01*				
E/0/0.15/54/7/15	Orange	$0.49 \pm 0.05*$	0.33±0.02*	0.11±0.01*	$0.0001 \pm 0.00*$	$0.83 \pm 0.04*$				
E/0/0.25/60/10/25	Vivid red	0.43±0.01*	0.31±0.02*	0.24 ± 0.02	$0.0001 \pm 0.00*$	0.93±0.02*				
E/3/0.15/30/3/10	Coral red	$0.94 \pm 0.07 *$	0.72 ± 0.07	0.21±0.04	0.0004 ± 0.00	1.80±0.13				
E/3/0.25/23/1/5	Coral red	$0.55 \pm 0.07*$	0.41±0.07*	0.28 ± 0.04	0.0003 ± 0.00	1.05±0.30*				
E/6/0.2/19/5/1	Vermilion	$0.46 \pm 0.04*$	0.31±0.04*	0.26 ± 0.06	0.0003 ± 0.00	$0.97 \pm 0.20*$				
G/30/11/2/1	Vermilion	0.53±0.06*	0.41±0.07*	0.31±0.02	0.0003 ± 0.00	1.08±0.26*				
		RN	M ₃ generation							
RE/0.15/2/2/15	Vivid red	1.15±0.09	0.79±0.06	$0.45 \pm 0.07*$	$0.0001 \pm 0.00*$	2.39±0.08				
RE/0.2/1/1/12	Carmine	$0.77 \pm 0.03*$	0.56±0.11	$0.50\pm0.08*$	$0.0002 \pm 0.00*$	1.84 ± 0.20				
RG/30/4/3/14	Carmine	$0.89 \pm 0.08 *$	0.70±0.16	$0.41 \pm 0.08*$	$0.0002 \pm 0.00*$	2.00±0.32				
RE/0.25/11/1/8	Vermilion	0.30±0.05*	0.24±0.03*	0.22 ± 0.02	$0.0001 \pm 0.00*$	0.76±0.08*				
*The values are significantly different than control at $p = 0.05$ according to the Student's t-test										

Every pigment, although, contributes to the colour of tissue; the resultant colour produced depends on the absolute content of a pigment and the relative content of each pigment in the tissue. To understand the role of latter, we used two metrics viz. the pigment contribution and pigment ratio. While pigment contribution conveys the proportion of a pigment among total betalain content in the tissue; the relative abundance of two pigments is understood from the pigment ratio. The pigment contribution of betalamic acid and betaxanthin was mostly affected

in the mutants. The pigment contribution of betalamic acid was increased in all the mutants induced through in vitro mutagenesis. Similarly, its pigment contribution was also increased in vermillion & vivid red along with coral red (E/3/0.25/23/1/5) & rose red (E/0/0.15/47/10/8) mutants induced through conventional mutagenesis. In the same manner the pigment contribution of betaxanthin was also increased in vermillion, coral red and rose red (E/0/0.15/47/10/8) mutants induced through conventional mutagenesis. In contrast in the vivid red mutant induced through in vitro mutagenesis the pigment contribution of betaxanthin was decreased. The pigment contribution of amaranthin was increased in the orange mutant, whereas it was decreased in carmine and vermillion mutants induced through in vitro mutagenesis (Table 4).

Mutant	Comb colour	Amaranthin	Betanin	Betalamic acid	Betaxanthin			
Control	Magenta	0.51	0.36	0.13	0.00010			
M ₃ generation								
E/0/0.15/47/8/5	Rose-red	0.57	0.39	0.12	0.00010			
E/0/0.15/47/10/8	Rose-red	0.56	0.40	0.26	0.00020			
E/0/0.15/54/7/15	Orange	0.59	0.40	0.13	0.00010			
E/0/0.25/60/10/25	Vivid red	0.46	0.17	0.26	0.00010			
E/3/0.15/30/3/10	Coral red	0.52	0.40	0.12	0.00020			
E/3/0.25/23/1/5	Coral red	0.52	0.42	0.27	0.00030			
E/6/0.2/19/5/1	Vermilion	0.47	0.32	0.27	0.00030			
G/30/11/2/1	Vermilion	0.49	0.38	0.29	0.00030			
		RM ₃ gene	ration					
RE/0.15/2/2/15	Vivid red	0.48	0.33	0.19	0.00004			
RE/0.2/1/1/12	Carmine	0.42	0.30	0.27	0.00010			
RG/30/4/3/14	Carmine	0.45	0.35	0.21	0.00010			
RE/0.25/11/1/8	Vermilion	0.39	0.32	0.29	0.00010			

Unlike pigment contribution, the pigment ratio was affected in all the mutants. The ratio of amaranthin : betanin was mostly reduced in the mutants. The same was true for the ratio of amaranthin : betalamic acid and amaranthin : betaxanthin. In the same way, the ratio of betanin : betalamic acid was decreased in all the mutants induced through in vitro mutagenesis. However, the ratio of betanin : betalamic acid was variedly affected in the mutants induced through conventional mutagenesis. The betanin : betalamic acid ratio was decreased in rose red (E/0/0.15/47/10/8), vivid red, coral red (E/3/0.25/23/1/5) and vermillion mutants. In contrast,

rose red (E/0/0.15/47/8/5), orange and coral red (E/3/0.15/30/3/10) had increased betanin : betalamic acid ratio. Notably, the betanin : betaxanthin ratio was decreased in coral red and vermillion mutants induced through conventional mutagenesis. Contrastingly, the betanin : betaxanthin ratio was increased in the orange and vivid red mutants. Similarly, the betalamic acid : betaxanthin ratio was increased in all the mutants induced through in vitro mutagenesis and the vivid red mutant (E/0/0.25/60/10/25). However, the coral red mutant (E/3/0.15/30/3/10) had strikingly decreased betalamic acid : betaxanthin ratio (Table 5).

Mutant	Comb colour	Am:Be	Am:Ba	Am:Bx	Be:Ba	Be:Bx	Ba:Bx		
Control	Magenta	1.41	3.96	3566.67	2.81	2533.33	900.00		
M ₃ generation									
E/0/0.15/47/8/5	Rose-red	1.46	4.78	4300.00	3.28	2950.00	900.00		
E/0/0.15/47/10/8	Rose-red	1.58	2.17	2600.00	1.38	1650.00	1200.00		
E/0/0.15/54/7/15	Orange	1.48	4.45	4900.00	3.00	3300.00	1100.00		
E/0/0.25/60/10/25	Vivid red	1.39	1.79	4300.00	1.29	3100.00	2400.00		
E/3/0.15/30/3/10	Coral red	1.31	4.48	2350.00	3.43	1800.00	525.00		
E/3/0.25/23/1/5	Coral red	1.34	1.96	1833.33	1.46	1366.67	933.33		
E/6/0.2/19/5/1	Vermilion	1.48	1.77	1533.33	1.19	1033.33	866.67		
G/30/11/2/1	Vermilion	1.29	1.71	1766.67	1.32	1366.67	1033.33		
		R	M ₃ generatio	n					
RE/0.15/2/2/15	Vivid red	1.46	2.56	11500.00	1.76	7900.00	4500.00		
RE/0.2/1/1/12	Carmine	1.38	1.54	3850.00	1.12	2800.00	2500.00		
RG/30/4/3/14	Carmine	1.27	2.17	4450.00	1.71	3500.00	2050.00		
RE/0.25/11/1/8	Vermilion	1.25	1.36	3000.00	1.09	2400.00	2200.00		
Am: Amaranthin, Be: Betanin, Ba: Betalamic acid, Bx: Betaxanthin									

Table 5. Pigment ratio in the comb of comb-colour mutants

Discussion

Isolation of flower/inflorescence colour mutants through conventional mutagenesis has been reported by Gnanamurthy and Dhanvel (2014) in *Vigna unguiculata* and Kumari and Kumar (2020) in gladiolous. Similarly, Latado et al. (2004) in *Chrysanthemum*, Bala and Singh (2015) in rose and Yoosumran et al. (2018) in *Dendranthemum grandiflora* have isolated colour mutants through in vitro mutagenesis. Anthocyanins impart colour to the flower/inflorescence in most Angiosperms (unlike cockscomb). Apart from this, other molecules such as flavonoids and carotenoids also play a role in imparting colour to the plant tissue. The alteration in the biosynthesis of these molecules might alter the colour of the tissue.

content of betalain pigments, which was evident in the present investigation, might be responsible for the comb-colour variation in the mutants. Moreover, all the pigments were not affected in same proportion in these mutants, which is evident from the variation in their pigment contribution and pigment ratio. Thus, apart from absolute content of the pigment; the relative content of each pigment also has played a role in determining the comb-colour of the mutants in cockscomb.

The biosynthesis of betalain pigments in plants begins from L-DOPA, which is converted to various molecules classified as betalain in a step-wise manner with each step being catalysed by an enzyme. As mentioned earlier, change in comb-colour of the mutants clearly indicates alteration in the content of betalain pigments in the cells. This proposition is substantiated by the TLC profile and quantification of a few betalain pigments in mutants in the present investigation. Onozaki et al. (1999) and Kobayashi et al. (2001) have opined that the mutation in a single gene result in the accumulation of the intermediates, which changes the flower colour. The synthesis of betacyanins and betaxanthins diverge from betalamic acid during the biosynthesis of betalains. While spontaneous condensation of betalamic acid with amino acids/amines synthesises betaxanthins; betacyanins are synthesised by the condensation of betalamic acid with cyclo-DOPA, which itself is derived from L-DOPA (Harris et al. 2012). Thus, there are several candidate enzymes, whose activity might influence the colour of comb. Mutation in any such gene could be responsible for induction of comb-colour in the present investigation. It is noteworthy, that pigment contribution of betalamic acid and betaxanthin was increased in most comb-colour mutants. This indicates comparative increase in the biosynthesis of betalamic acid and betaxanthin among all the pigments. Betalamic acid is derived from 4,5seco-DOPA, which is formed by cleavage of the ring of L-DOPA. Similarly, L-DOPA is also the precursor of cyclo-DOPA. As mentioned above condensation of betalamic acid with amino acids/amines and cyclo-DOPA forms betaxanthins and betacyanins, respectively. The increase in pigment contribution of betalamic acid indicates that channelizing the L-DOPA for biosynthesis of 4,5-seco-DOPA is favoured over the biosynthesis of cyclo-DOPA. Moreover, increase in the pigment contribution of betaxanthin in most mutants indicates that condensation of betalamic acid with amino acids/amines is preferred over its condensation with cyclo-DOPA, which presumably is in lesser amount in the mutants. Overall, this might have reduced the pigment ratio of betacyanins-amaranthine and betanin-in the mutants. Whatsoever, the absolute content of all the molecules quantified is decreased in the mutants. This might be due to reduction in either the content of L-DOPA or its downstream metabolism. Therefore, it is likely that the gene involved in these reactions might be affected in the mutants.

Notwithstanding anything said above, apart from the pigments analysed in comb-colour mutants, there might be other pigments in the mutants which were affected and contributed to the change in comb-colour. This is reasonable from the fact that neither the absolute pigment content nor the relative pigment content of any two mutants with same comb-colour was similar, if not identical, in the present investigation.

Conclusion

The comb-colour mutants isolated in the present investigation can serve two purposes. First, these mutants can be utilised in crop improvement and second, they can be a system to study betalain biosynthesis in plants.

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